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# Macro-Hybrid and Micro-Hybrid of Fish

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## ABSTRACT

This study reviews the progress in fish breeding, focusing on distant hybridization and gynogenesis, and introduces the theory of macro-hybrid and micro-hybrid. The macro-hybrid refers to the allo-progeny derived from distant hybridization, including the allo-progenies. The micro-hybrid refers to the auto-progenies derived from distant hybridization or heterologous sperm-induced gynogenesis, which possess a genome derived predominantly from the maternal but contain DNA fragments originating from the paternal. Macro-hybrids induce significant phenotypic changes, while micro-hybrids improve growth rate and stress resistance. Both hybridization and gynogenesis exhibit high selective pressure. This review elucidates how selective pressures (homozygosity + heterologous sperm + cold/heat shock) of the heterologous sperm-induced gynogenesis contribute to the generation of effective genetic variations. The study also presents the technologies of macro-hybrid and micro-hybrid. In macro-hybrids of distant hybridization, parental chromosome numbers are closely matched, enabling the formation of fertile allo-diploid and allo-tetraploid strains. In micro-hybrid of distant hybridization, equal or differing chromosome numbers between parents yield fertile auto-tetraploid and auto-diploid strains. In micro-hybrid of heterologous sperm-induced gynogenesis, equal or different chromosome numbers between parents yield auto-diploid strains. The integration of heterologous sperm-induced gynogenesis, back-cross, and self-cross strategies can address the issue of all-female progeny resulting from gynogenesis. A series of auxiliary breeding techniques is established to support the macro-hybrid and micro-hybrid breeding. Case studies of superior fish strains developed through macro-hybrid and micro-hybrid breeding are presented. The establishment of macro-hybrid and micro-hybrid theory and breeding technologies holds significant value for fish breeding.

## 1 | Overview of Breeding Techniques in China and Abroad

Aquatic products provide approximately one-third of the high-quality animal protein consumed by the Chinese population. Among them, fish account for 42% of aquatic products, making

them the largest category and a vital source of protein [1, 2]. The seed industry contributes the most to the aquatic products value chain. Therefore, fish breeding techniques play a crucial role in aquaculture. The overall purpose of aquaculture breeding is to develop improved aquatic species with enhanced traits such as growth, disease resistance, and environmental adaptability to

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support sustainable and efficient production. The following is an overview and analysis of fish breeding techniques both in China and internationally:

### 1.1 | Selective Breeding

Selective breeding is one of the most commonly used breeding methods and holds an essential position in fish breeding [3]. Its primary goal is to identify and select individuals or populations with desirable traits from one or more groups. Traditional selective breeding approaches in fish include mass selection, family selection, parent selection, and combined selection [4–6], as well as BLUP (Best Linear Unbiased Prediction) based on estimations of breeding values [7–9]. These techniques have been applied in the selective breeding of various freshwater [10–12] and marine fish species [13–16]. Moreover, molecular marker-assisted selection (MAS) [17–32], genomic selection (GS) [33–41], and genome-wide association studies (GWAS) [42–47] have also been applied in fish breeding. Conventional selective breeding is simple, inexpensive, and has been widely applied to improve growth, yield, and adaptation by utilizing natural variation. However, it is slow, imprecise, and limited by available genetic diversity, with risks of inbreeding and trade-offs [48, 49]. MAS enhances precision by targeting specific genes or QTLs associated with important traits, accelerating genetic gain and reducing environmental bias. Yet, MAS is most effective for traits controlled by few major genes and may have limited impact on complex, polygenic traits [50–52]. GS overcomes this limitation by using genome-wide markers to predict breeding values, enabling faster, more accurate selection for both simple and complex traits. Although GS requires significant investment in genotyping, bioinformatics, and training populations, it provides the highest efficiency and long-term genetic improvement compared to conventional and MAS approaches [53, 54].

### 1.2 | Germ Cell Transplantation and Nuclear Transfer Breeding

Nuclear transfer has been applied to the development of nucleocytoplasmic hybrid fish and the generation of homozygous diploid fish. Experiments in teleost species, such as the goldfish (*Carassius auratus*) and the Chinese bitterling (*Rhodeus sinensis*), have demonstrated that blastula-stage nuclei possess totipotency, capable of directing enucleated eggs to develop into embryos and adults. For example, by transplanting the kidney cell nucleus of triploid crucian carp (*Carassius auratus*) into enucleated diploid crucian carp eggs, researchers successfully produced fertile cloned fish, confirming the genetic and developmental totipotency of differentiated fish somatic nuclei for the first time [55]. However, low survival rates in nucleocytoplasmic hybrid fish remain a challenge and require further research [56].

Researchers have also transplanted primordial germ cells from rainbow trout into the peritoneal cavity of newly hatched masu salmon fry, resulting in masu salmon producing rainbow trout offspring, demonstrating the feasibility of germ cell transplantation in fish [57, 58]. Subsequent studies successfully established spermatogonial stem cell lines (e.g., SG3) and haploid embryonic stem cell lines in species such as medaka and rainbow trout, creating germ cell transplantation systems and applying semi-clonal

techniques to produce fertile individuals [59–61]. Germ cell transplantation in fish can be classified into intra-species, inter-species, and triploid recipient transplantation. Among them, interspecific transplantation has shown that donor cells can undergo accelerated differentiation in heterologous recipients. Triploid recipients have also successfully produced purebred offspring, highlighting the mechanism and potential applications of environmentally regulated reproductive development [58, 62–64]. Both germ cell transplantation and nuclear transfer techniques continue to play important roles in fish breeding research [60, 63, 65–80]. While nuclear transplantation has demonstrated the totipotency of fish somatic cells, immune rejection of donor cells remains a significant obstacle in cross-species transplantation, significantly affecting chimeric survival and the efficiency of germ cell colonization [81]. Although germ cell transplantation transcends species barriers, the long-term reproductive viability of donor-derived gametes is questionable-heterologous microenvironments may induce epigenetic dysregulation or aberrant differentiation in germline stem cells [82]. From a technical perspective, challenges in achieving precise microsurgical manipulation and synchronizing recipient embryos represent major procedural bottlenecks, restricting large-scale implementation. Existing methodologies function mainly as proof-of-concept [83], while meaningful application in sustainable aquaculture requires advances in immunocompatibility, automation, and thorough long-term safety evaluation.

### 1.3 | Physical and Chemical Induction of Polyploidy

Artificial polyploid breeding can be achieved through physical or chemical induction. Physical methods primarily involve heat or cold shock and hydrostatic pressure to induce triploid or tetraploid individuals [84–90], while chemical induction methods use agents such as colchicine to achieve the same results [91–95]. These approaches differ from the polyploid populations obtained via distant hybridization. To date, no reports have successfully established bisexual fertile tetraploid fish lines using these methods, possibly due to difficulties in achieving exact tetraploid chromosome numbers.

We further posit that while physical or chemical polyploid induction can achieve high success rates, they carry significant limitations, including challenges in establishing stably fertile tetraploid lines [93], residual chemical inducers (e.g., colchicine), and elevated teratogenic risks from physical stressors. At the same time, escaped sterile individuals may compete with wild conspecifics, whereas fertile tetraploid escapees pose a risk of genetic pollution through introgression [96]. Currently, these techniques are primarily used in experimental settings. For their broader application in scalable breeding, it is essential to integrate them with complementary strategies and to conduct systematic evaluations of their long-term health and ecological impacts.

### 1.4 | Transgenic and Gene Editing

The world's first transgenic crucian carp was produced in China [97], followed by several related studies [97–102]. The use of transgenes to introduce exogenous genes for the enhancement

of desirable traits has achieved notable success in multiple fish species [98, 103–107]. Aquaculture research primarily focuses on enhancing growth, disease resistance, cold tolerance, and exploring life mechanisms [98, 108–119]; however, concerns about the ecological risks of transgenic fish remain. The genetically modified Atlantic salmon, developed in the United States, became the world's first approved transgenic fish for commercial sale [120–122].

Gene editing enables direct manipulation of DNA and has been widely applied in model organisms such as zebrafish [123–132] and in economically important fish species [133–137]. Recently, the knockout of the *cntd1* gene enabled the production of unreduced gametes and polyploid zebrafish, offering new avenues for creating triploid and tetraploid fish [138]. Furthermore, gene editing has shown promising results in reducing intramuscular bones in fish [139–146]. Although significant progress has been made in transgenic and genome editing for aquaculture breeding, their systemic risks remain insufficiently evaluated. To prevent the pitfalls of unchecked technological optimism, it is essential to couple these advancements with thorough life-cycle ecological risk assessments, ethical oversight, and consideration of societal acceptance.

### 1.5 | Distant and Close Hybridization Breeding

Hybrid breeding not only produces heterosis but also creates progeny that combine superior traits from both parents, surpass parental traits, or exhibit novel characteristics. This method has become a widely applied, effective, and rapid approach to introduce genetic variation [147–158]. Through hybridization, advantageous genes from different parents can be combined, significantly improving the overall performance of the offspring [159–161]. As of 2024, hybridized fish varieties account for 32% of all new fish breeds approved in China, among which distant hybrids constitute 68%. Some of these new varieties have already shown excellent performance in commercial applications, such as *Channa argus* × *Channa maculata*, *Carassius auratus cuvieri* × *Carassius auratus red var.*, and *Megalobrama amblycephala* × *Culter alburnus* [162–164].

Since the first report on distant hybridization in fish in 1558, studies have been conducted on over 1080 fish species across 56 families worldwide, with a primary focus on orders such as Centrarchiformes [165, 166], Cypriniformes [167–172], Poeciliidae [173], and Salmonidae [174, 175]. Since the late 1950s, breeding research using hybridization has resulted in a series of high-quality fish varieties, promoting the development of aquaculture [176–183].

Our team has conducted extensive research on fish distant hybridization, overcoming the three major challenges: low survival rate, reproductive difficulties, and identification challenges. They have elucidated the main genetic and reproductive patterns in distant hybrids, established both one-step and multi-step breeding strategies, and successfully cultivated fertile tetraploid lines, naturally gynogenetic strains, and improved triploid fish varieties [155, 159, 184–188]. Among seven distant hybridization combinations, seven naturally gynogenetic fish types have been developed [149, 150, 153, 189–193].

Nonetheless, more precise breeding through distant hybridization remains a topic for further investigation. Building on previous work, our team has integrated distant hybridization with heterologous sperm-induced gynogenesis, establishing the concepts and technologies of macro-hybridization and micro-hybridization, thereby enhancing the accuracy and applicability of these breeding approaches [155]. While distant hybridization demonstrates significant efficacy in developing novel germplasm (e.g., allopolyploids, gynogenetic lines) and advancing commercial cultivar development (accounting for 32% of new aquatic varieties in China), its potential risks remain systematically unassessed. Hybrid progeny frequently exhibit genetic instability leading to trait segregation [194], and large-scale adoption exacerbates genetic homogenization within farmed populations. Escaped hybrids may compromise the genetic integrity of wild relatives through introgression [195]. Current regulatory frameworks lack sufficient ecological risk assessment for the environmental release of hybrids, with reproductive barriers in inter-familial/genus crosses (e.g., fertility fluctuations, offspring deformities) persisting as technical constraints. Integrating genomic surveillance to quantify introgression risks and establishing an interdisciplinary evaluation framework are imperative to reconcile breeding benefits with ecological sustainability [196].

### 1.6 | Gynogenetic (and Androgenetic) Breeding

Gynogenesis can be classified into artificial and natural types. Artificial gynogenesis is often achieved through heterologous sperm-induced activation. In this process, eggs are activated by sperm from a different species, but the resulting offspring inherit genetic material almost exclusively from the maternal genome, leading to highly homozygous populations. Cold (or heat) shock treatment is applied to prevent the extrusion of the second polar body and achieve chromosome doubling.

Since the 1960s, scientists worldwide have successfully induced gynogenesis and androgenesis in various fish species [197–207].

Over the years, we have developed multiple superior artificially gynogenetic fish [208–217] and naturally gynogenetic strains [184, 218–221], many of which exhibit excellent or unique traits and constitute novel germplasm resources. For instance, gynogenetic mirgial carp exhibit enhanced cold tolerance [210], gynogenetic mandarin fish exhibit accelerated embryonic development [208], gynogenetic bluntnose black bream show increased hypoxia tolerance [214, 222], and gynogenetic northern snakehead shows faster growth [217]. Additionally, gynogenetic white crucian carp demonstrate improved disease resistance and growth [212], and gynogenetic koi exhibit more vivid coloration [223]. Naturally gynogenetic grass carp exhibit higher amino acid content in muscle tissues [191], while naturally gynogenetic bluntnose black bream exhibit superior nutrition and muscle characteristics [224].

Androgenesis refers to the development of embryos initiated by inactivated eggs fertilized by normal sperm, with the paternal genome guiding development. Although rare in nature, cases of androgenesis have occasionally been observed in fish hybrids [225–227] (for example, carp and grass carp hybrid, white crucian carp, and bream hybrid). Artificial

induction of androgenesis has been successfully achieved in several species [228–232]. Notably, the use of diploid sperm from tetraploid fish can bypass the need for chromosome doubling treatments and significantly improve success rates [159, 204, 205, 233–235].

For instance: In Cypriniformes with buoyant eggs (e.g., grass carp [236], silver carp, bighead carp, mrigal carp [210]), the optimal cold shock duration is 10–15 min; for adhesive eggs (e.g., carp, crucian [212], blunt snout bream [214], culter), it is 20–40 min; for Perciformes (e.g., largemouth bass [211], mandarin fish [208], northern snakehead [217]) and Siluriformes (e.g., channel catfish, native catfish, yellow catfish), the ideal cold shock duration is 15–20 min; empirical evidence from extensive experimental results underscores the necessity of establishing protocols with quantifiable survival rates. Although androgenesis/gynogenesis has successfully generated diverse elite lines, its application faces persistent bottlenecks: artificially induced survival rates typically fall below 5%, and ploidy manipulation techniques frequently induce teratogenic effects. These inherent limitations represent critical priority areas for future optimization.

## 2 | Comparison of Breeding Techniques

From 1996 to 2024, a total of 306 new aquaculture varieties were approved in China. Among them, 43% were developed through selective breeding, 35% through hybridization (distant and close), and 5% through gynogenesis. Techniques such as germ cell transplantation, nuclear transfer, transgenics, and gene editing have yet to be reflected in newly approved varieties, mainly due to regulatory and technical constraints [237].

Selective breeding, hybridization, and gynogenesis have distinct characteristics. From a phenotypic perspective, offspring derived from selective breeding or gynogenesis generally resemble the paternal and maternal phenotypes (especially in heterologous sperm-induced gynogenesis). Genetically, these two methods also result in relatively minor changes. In contrast, distant hybridization tends to induce greater changes in both phenotype and genetic composition. This comparison provides a framework for selecting breeding methods: for minimal genetic and phenotypic alteration, selective breeding and gynogenesis are appropriate; for significant innovation in phenotype and genetics, distant hybridization is preferable.

Furthermore, selective pressure plays a significant role in both distant hybridization and gynogenesis. For example, in heterologous sperm-induced gynogenesis, low survival rates due to strong selection pressure act as a powerful screening mechanism [238]. Thus, in addition to its inherent genetic effects, heterologous sperm-induced gynogenesis also exhibits selection effects. Selective breeding, therefore, can either be applied independently or integrated with other breeding techniques for enhanced outcomes.

While selective breeding and gynogenesis maintain phenotypic stability, they neglect the adaptive decline caused by inbreeding depression and the fixation of deleterious alleles [239, 240]. Although the innovative potential of distant hybridization is

often highlighted, insufficient attention is given to its inherent reproductive barriers (e.g., hybrid sterility), trait segregation, and ecological risks (e.g., genetic pollution from escaped hybrids). Claims regarding the “broad applicability” of heterologous sperm-induced gynogenesis lack empirical support—survival rates below 5% and high deformity rates across multiple species challenge its technical universality [241]. Crucially, the synergistic potential between modern biotechnologies (e.g., gene editing, germ cell transplantation) and traditional methods remains systematically overlooked. Future breeding strategies should integrate these complementary approaches rather than relying on a single technique.

## 3 | Theory and Technologies of Macro-Hybrid and Micro-Hybrid

We present the theory and technologies of macro-hybrid and micro-hybrid systems, based on extensive practical experience with distant hybridization and gynogenesis.

**Macro-Hybrid:** The macro-hybrid refers to the allo-progenies derived from distant hybridization, including the allo-diploid and allo-tetraploid progenies (Figure 1). In macro-hybrids of distant hybridization, parental chromosome numbers are closely matched, enabling the formation of fertile allo-diploid and allo-tetraploid strains. In micro-hybrid breeding of distant hybridization, equal or different chromosome numbers between parents yield fertile auto-tetraploid or auto-diploid strains (Figure 1).

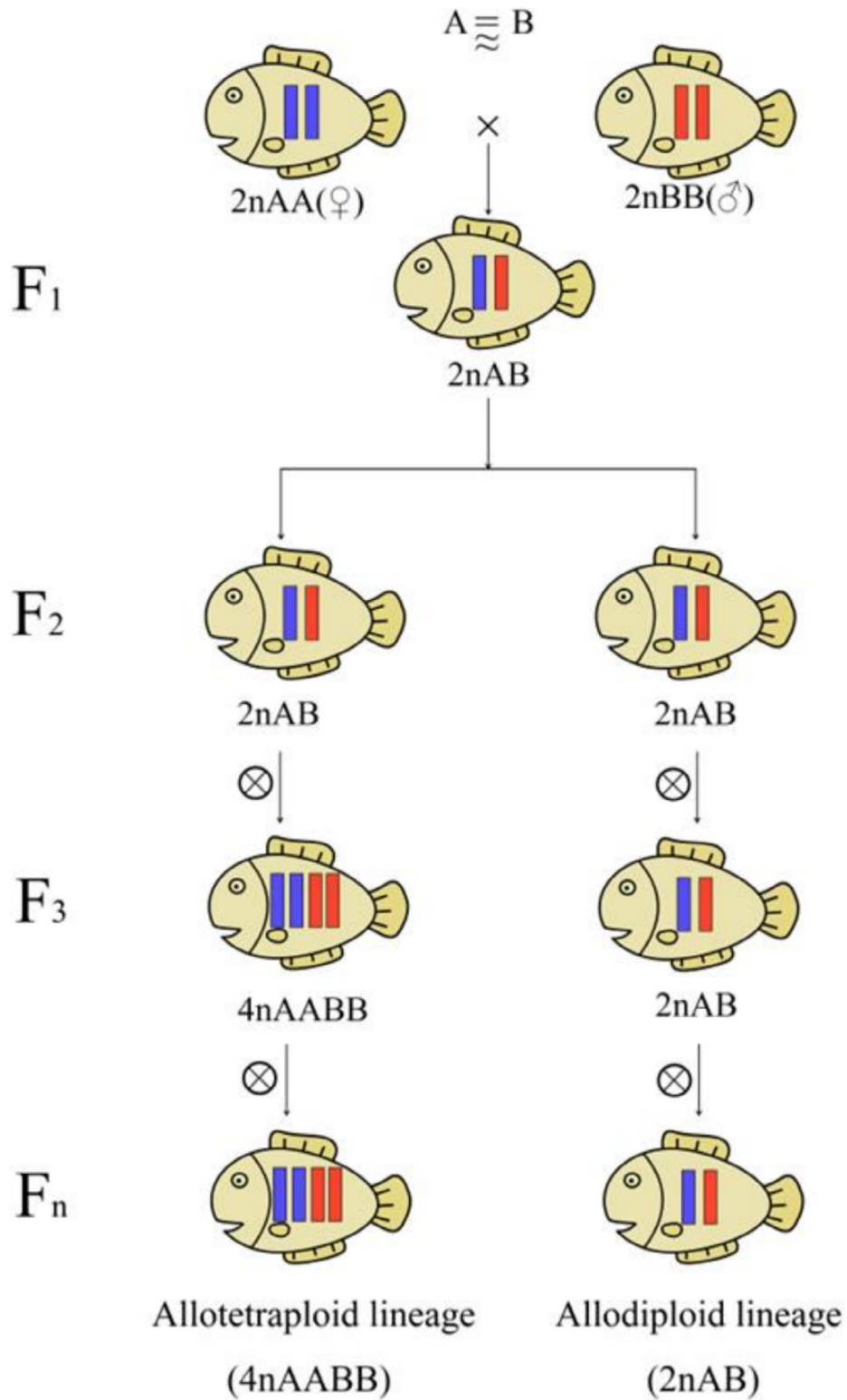
**Micro-Hybrid:** The micro-hybrid refers to the auto-progenies derived from distant hybridization or heterologous sperm-induced gynogenesis, which possess the genome from the maternal parent predominantly but contain DNA fragments originating from the paternal (Figure 2a,b). In the micro-hybrid of distant hybridization, equal or different chromosome numbers between parents yield fertile auto-tetraploid or auto-diploid strains. In the micro-hybrid of heterologous sperm-induced gynogenesis, equal or different chromosome numbers between parents yield auto-diploid strains. The integration of heterologous sperm-induced gynogenesis, back-cross, and self-cross strategies can address the issue of all-female progeny resulting from gynogenesis when the maternal parent possesses an XX sex determination system, thereby enabling rapid expansion of the breeding population (Figure 3). Amplification of the micro-hybrid effect was achieved through repeated crossings of multiple populations that already possessed the effect (Figure 2c).

## 4 | Classifying Macro-Hybrid and Micro-Hybrid Lineages in Fish Breeding

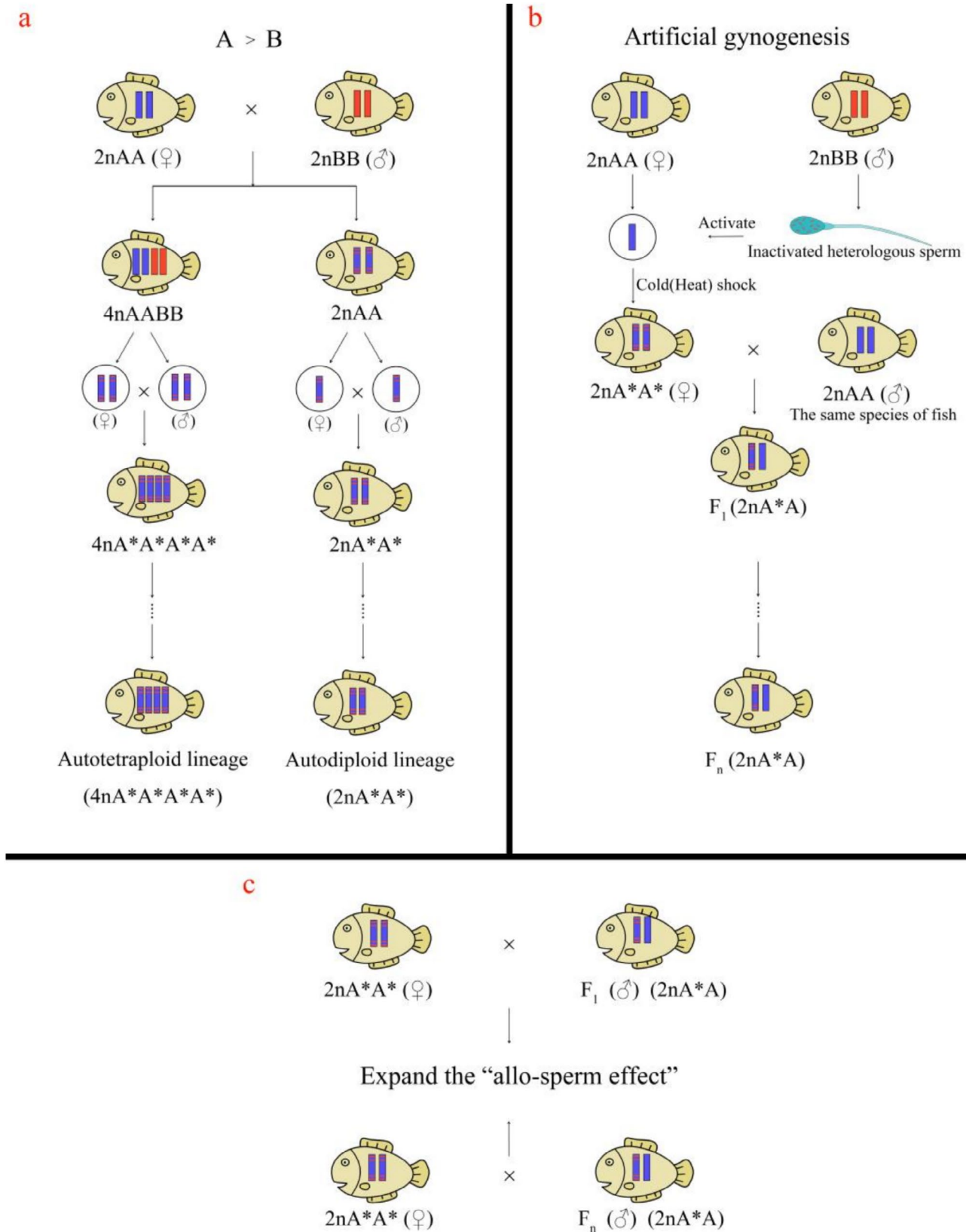
Based on extensive studies of distant hybridization and gynecetic development in fish, we systematically compared the similarities and differences between these two processes. It was found that both distant hybridization and gynecetic development involve the fertilization of eggs by heterologous sperm and the initiation of embryonic development, thereby sharing the fundamental feature of “hybridization.” The difference lies in the specific manipulations employed: gynecetic development involves sperm inactivation and special treatment of fertilized



# Macro-hybrid Breeding Workflow



**FIGURE 1** | Schematic diagram of macro-hybrid breeding technologies. The symbol “ $\approx$ ” indicates that the chromosome numbers of the two groups are numerically similar, differing by no more than two chromosome sets ( $2n$ ), and are considered to have broadly compatible genomes for the purpose of hybridization.



**FIGURE 2** | Schematic diagram of micro-hybrid breeding technology (The self-fertilization of  $4nAABB$  has been consistently demonstrated to produce exclusively  $4nAAAA$  offspring in multiple experiments). (a) The auto-progenies derived from distant hybridization, which possess the genome from the maternal dominantly but contains DNA fragments originating from the paternal; (b) The auto-progenies derived from heterologous sperm-induced gynogenesis, which possess the genome from the maternal dominantly but contains DNA fragments originating from the paternal; (c) Amplification of the micro-hybrid effect was achieved through repeated crossings of multiple populations that already possessed the effect.

eggs (such as cold or heat shock to promote chromosome duplication), whereas distant hybridization does not involve such interventions (Figure 4) [155].

We proposed the theoretical framework of “macro-hybrid” and “micro-hybrid”. Distant hybridization can yield both macro-hybrid and micro-hybrid lineages, whereas gynogenesis yields only micro-hybrid lineages.

We conducted systematic studies on the genetic patterns and mechanisms underlying trait formation in fish derived from distant hybridization and gynogenesis. Comparative analyzes at the gene, transcriptome, and genome levels were performed between various fish lineages, including allo-diploid BSB-culter hybrids, culter-BSB hybrids, and allo-tetraploid crucian-carp hybrids and their respective parents. These studies revealed the co-existence and stable inheritance of biparental subgenomes. For instance, both the allo-tetraploid AABB and the allo-diploid AB individuals maintain two coexisting subgenomes, which can be

stably passed on. Furthermore, recombination between homologous sequences of different subgenomes was observed, and this recombination was found to accumulate over successive generations [242, 243]. The expression of recombinant genes between subgenomes significantly influences the genetic characteristics and trait development of hybrid fish [244, 245].

Moreover, analyzes of auto-tetraploid crucian and auto-tetraploid carp lineages (originating from distant hybridization) and their parental species revealed that the maternal genome predominates (e.g., tetraploid AAAA and diploid AA). Insertion of paternal DNA fragments was also observed in these genomes [246, 247]. Similar insertions were detected in various fish derived from artificial gynogenesis [155, 208, 217].

Macro-hybrids are typically associated with prominent morphological changes, whereas micro-hybrids tend to exhibit subtler morphological differences but significant improvements in growth rate, stress resistance, and other traits.

The components of heterologous sperm-induced gynogenesis, including homozygosity, heterologous sperm, and cold (heat) shock, are all associated with intense selective pressure. The “homozygosity effect” arises from the chromosomal doubling of the maternal genome, which purges deleterious recessive alleles through homozygous expression (e.g., lethality or disease), while retaining and accumulating beneficial mutations. The heterologous sperm effect constitutes a form of micro-hybrid variation that may also eliminate unfavorable traits and preserve advantageous ones. Thermal shock eliminates weaker individuals, contributing further to selection. The combination of these three factors including homozygosity, heterologous sperm, and thermal shock-creates intense selective pressure that identifies and enriches for superior individuals or populations. Earlier research suggested that the homozygosity effect of gynogenetic development was equivalent

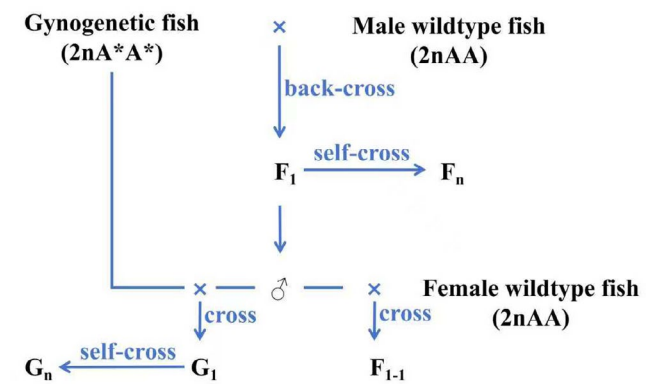


FIGURE 3 | Breeding strategies of micro-hybrid.

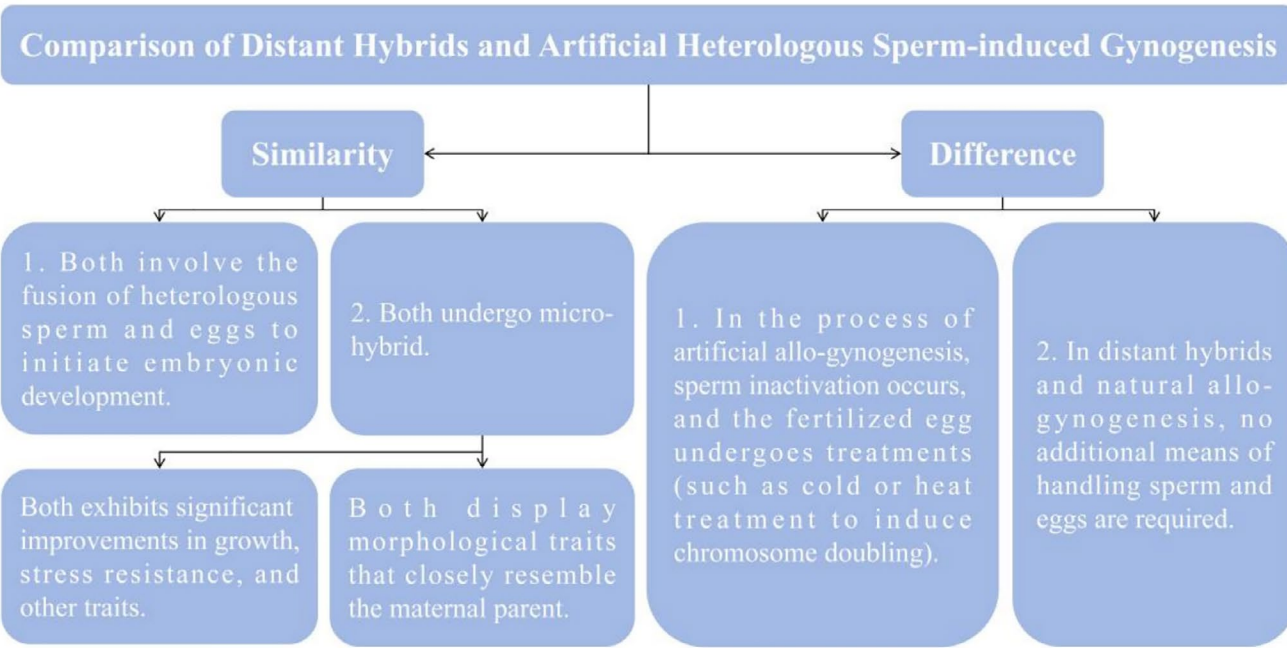


FIGURE 4 | Comparison of distant hybridization and artificial heterologous sperm-induced gynogenesis.

to 8–10 generations of inbreeding [248, 249]. We propose that the combined selective effects of homozygosity, heterologous sperm, and cold (heat) shock likewise approximate 8–10 generations of selection, establishing a new theoretical framework.

Both the homozygosity effect and the heterologous sperm effect are potent sources of genetic variation in fish breeding. These findings underscore the principle that macro-hybrids result in notable phenotypic changes, while micro-hybrids lead to subtler phenotypic shifts [155, 217]. Our team provides substantial evidence supporting the heritability of micro-hybrid effects in gynogenetic fish through back-cross and self-cross [250], thereby laying a theoretical foundation for developing gynogenetic breeding strategies based on micro-hybridization.

Traditionally, gynogenesis has been regarded primarily as a method for producing highly homozygous populations. Few have recognized that homozygosity itself represents a form of genetic variation. Our team argues that the essence of breeding lies in identifying variation and seeking useful individuals or populations within diverse genetic backgrounds. Thus, increased homozygosity is a form of beneficial variation. Moreover, we highlight that sperm inactivation and cold (heat) shock during gynogenesis are critical drivers of variation. Coupled with the theory of micro-hybrid, these insights provide a more comprehensive understanding of the principles underlying gynogenetic breeding and support the development and broader application of relevant breeding techniques.

Based on survival rates (from fertilized egg to fry) in experiments using cohorts of 2000 eggs, the rates in increasing order were [208–211, 214, 217]: mandarin fish: 0.01%–0.02%; Mrigal carp: 0.5%; northern snakehead: 1%–5%; goldfish, *Hemibarbus maculatus*: 2%–8%; blunt snout bream: 1%–3%; tetraploid lineages: 2%–5%; common carp and koi: 3%–8%; red crucian carp and white crucian carp: 5%–9%; largemouth bass: 6%; grass carp and large yellow croaker (collaborating with Professor Xinhua Chen's team at Fujian Agriculture and Forestry University): 10%. These rates, ranging from 0.01% to 10%, demonstrate the low survival rates and intense selection pressures associated with heterologous sperm-induced gynogenesis. The few surviving individuals typically exhibit enhanced resistance to stress and disease, as well as accelerated growth.

In the field of gynogenesis, our team made significant theoretical and technological innovations by emphasizing the mutational effects of both homozygosity and heterologous sperm. The integration of distant hybridization and gynogenesis led to the proposal of the macro-hybrid and micro-hybrid framework, which forms a solid foundation for the establishment and application of corresponding breeding technologies. Given the strong selective effects inherent in heterologous sperm-induced gynogenesis, this micro-hybrid breeding approach can be applied not only to natural fish and fertile distant hybrids but also to their descendants. We have recently documented the successful induction of gynogenesis in marine bivalves. This suggests the potential for broad adaptability of these technologies across diverse species. While numerous global cases of gynogenesis and hybrid fish exist, few have yielded established breeding lines. Our macro- and micro-hybridization approaches provide a methodological framework to guide subsequent research in this domain.

## 5 | Applications of Macro-Hybrid and Micro-Hybrid

Based on extensive and systematic practical and theoretical studies of fish distant hybridization and gynogenetic development, our team proposed the theories of macro-hybrid and micro-hybrid and subsequently designed and established corresponding breeding technologies. These approaches build upon previously established “one-step” and “multi-step” breeding methods [188, 246, 251], further refining them and expanding their application scope. The contents of the breeding technologies are detailed below:

**Macro-Hybrid Breeding** (Figure 2) involves designing crosses between fish species with equal chromosome numbers. Their offspring may form fertile allo-tetraploid or allo-diploid fish, which can then self-cross to establish respective allo-tetraploid or allo-diploid lines, each containing both parental subgenomes. This method effectively addresses survival issues (via chromosomal compatibility) and reproductive challenges (by enabling both meiotic and ameiotic gamete formation). Using this strategy, multiple tetraploid and diploid fish lines have been developed [149, 151, 156, 159, 164, 220, 245, 252, 253], creating new germplasm resources.

**Micro-Hybrid Breeding** (Figure 2) is designed using specific parental combinations and breeding strategies based on both distant hybridization and gynogenesis. In distant hybridization, the parents are chosen such that the maternal species has significantly more chromosomes than the paternal species. This enables the formation of fertile auto-tetraploid and auto-diploid offspring dominated by the maternal genome but containing inserted paternal DNA fragments. These offspring can self-cross to form stable auto-tetraploid and auto-diploid lines. This strategy effectively addresses survival and reproductive barriers, yielding valuable germplasm resources [153, 161, 192, 218, 219, 221, 227, 254, 255].

In heterologous sperm-induced gynogenesis, parental species from different taxa are selected. The primary generation produced is typically a fertile diploid female (if the sex-determination system is homogametic XX), dominated by the maternal genome and with inserted paternal DNA fragments. These individuals are then crossed with ordinary males to generate the first filial generation, which is subsequently self-crossed to produce the second generation. Male individuals from either generation can be back-crossed with original gynogenetic females to further enhance the micro-hybrid effect (Figure 2). This method overcomes the lack of males and the limited original population size in gynogenetic offspring.

In natural gynogenetic offspring resulting from distant hybridization, both sexes may be present, and the micro-hybrid effect is observable in both. This effect can be inherited through self-crossing. In addition to self-cross, back-cross with normal males or females is also proposed as a breeding strategy to propagate the micro-hybrid effect. During gynogenesis, paternal DNA segments may integrate via special mechanisms resembling distant hybridization, generating sexually dimorphic populations. These male and female individuals can then be used in a self- or back-cross to expand the population. This represents a



novel fusion of gynogenetic and distant hybridization processes. Notably, reducing sperm vitality may enhance offspring survival in certain cases. Partial inactivation, non-inactivation, or complete inactivation of sperm during gynogenesis can lead to diverse outcomes, a novel concept not explored in prior studies.

Given the high selection intensity, low retention rates, and small initial populations in heterologous sperm-induced gynogenesis, the establishment of the micro-hybrid theory and breeding technologies effectively overcomes these limitations, enabling rapid expansion of effective populations. The core of this system lies in clearly distinguishing high-quality germ-plasm types: first, by obtaining primary populations through gynogenesis; then expanding these using self- or back-cross. Although the micro-hybrid effect may be diluted in later generations, experimental evidence demonstrates that advantageous traits in the original gynogenetic fish can be stably inherited and expressed through these means [250], greatly enhancing breeding efficiency [155].

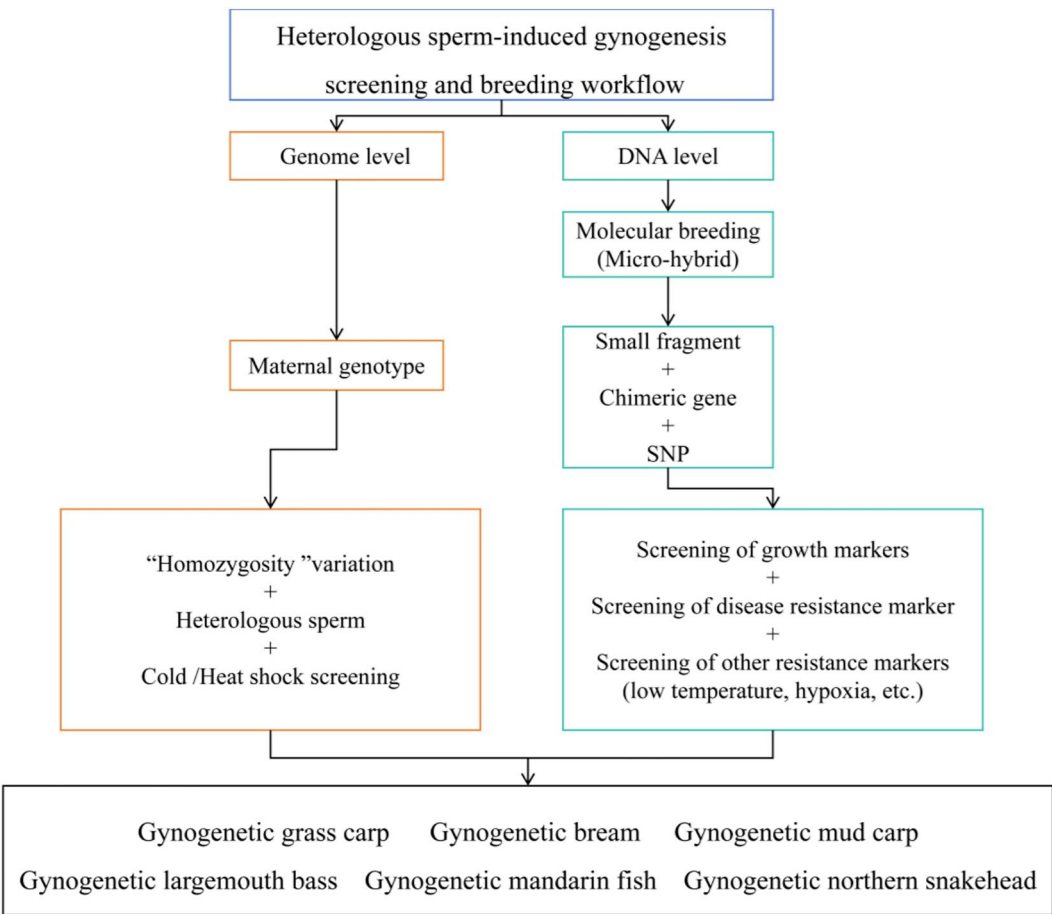
Macro-hybrid and micro-hybrid provide a strong theoretical and technical foundation for the previously established one-step and multi-step breeding methods [155, 188], while enhancing the precision of the multi-step breeding method. For instance, one-step breeding emphasizes the use of  $F_1$  distant hybrids with equal parental chromosome numbers, while macro-hybrid breeding highlights the biparental subgenome composition of these  $F_1$  hybrids. Multi-step breeding

emphasizes the fertility of distant hybrid offspring and their role as new breeding parents; macro-hybrid and micro-hybrid further elucidate the genomic compositions of the derived allo- and auto-tetraploid and diploid lines. Moreover, the breeding technologies organically integrate distant hybridization and gynogenesis, significantly broadening the application scope of these approaches.

Gynogenetic breeding techniques reflect both genetic variation and selection mechanisms at the chromosomal (genomic) and DNA levels. These processes include genomic homozygosity, micro-hybrid effects, and cold (heat) shock-induced selection. Together, they improve growth rate, disease resistance, cold tolerance, and hypoxia tolerance in gynogenetic offspring (Figure 5).

**TABLE 1** | Micro-level auxiliary breeding techniques.

Micro-level auxiliary breeding techniques	
1	Genome sequencing is used for overall subgenomic identification
2	Molecular marker screening, employed for partial subgenome recognition
3	PCR, used for identifying paternal DNA fragments
4	SNP site detection and related genotyping tools



**FIGURE 5** | Combination of heterologous sperm-induced gynogenesis and molecular screening.

Compared with distant hybridization, gynogenesis is less restricted by parent compatibility and thus can be more widely applied across fish species. However, it is generally more difficult to obtain fertile tetraploids through gynogenesis alone, something that distant hybridization can achieve more effectively.

In terms of breeding guidance, a macro-level design perspective can be adopted based on the fact that distant hybridization may result in two types of effects: macro-hybrid and micro-hybrid.

**TABLE 2** | Macro-level auxiliary breeding techniques.

Macro-level auxiliary breeding techniques	
1	Compatibility between sperm head size and egg micropyle [256]
2	Genetic relationship and trait complementarity between parental species, ensuring the survivability and trait contribution of the paternal genome
3	Cryopreservation of sperm
4	Optimization of the duration of sperm inactivation
5	Prolonging the growth period of hybrid fish to ensure sexual maturity
6	Pooling sperm from low-fertility males to ensure fertilization success, designing parental crosses based on chromosomal inheritance
7	Supporting techniques such as flow cytometry, chromosome analysis, FISH, and gamete assessment
8	Drip incubation and temperature-controlled hatching

Additionally, heterologous-sperm-induced gynogenesis can generate micro-hybrid effects. Macro-hybrids are typically characterized by substantial changes in both phenotype and genetic composition, whereas micro-hybrids exhibit relatively minor alterations in these aspects. These distinctions provide a theoretical framework for the purposeful and precise design and implementation of breeding strategies.

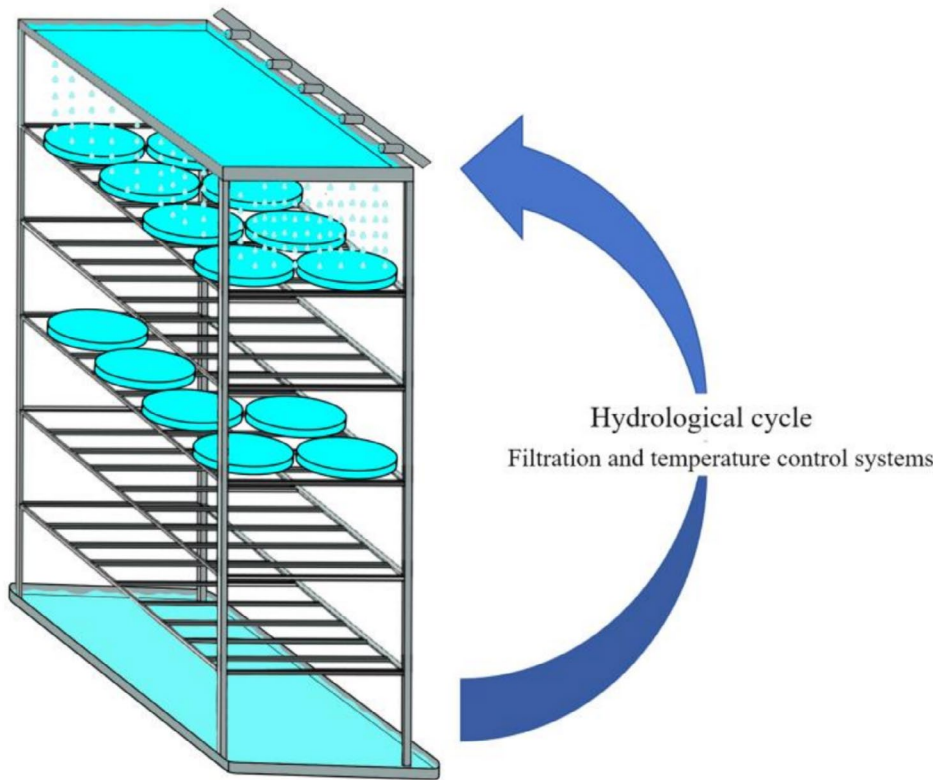
6 | Auxiliary Breeding Techniques

Centering on macro-hybrid and micro-hybrid, we developed a comprehensive breeding system by incorporating various auxiliary techniques. These include both micro-level and macro-level auxiliary breeding approaches (Tables 1 and 2).

These techniques have been applied to identify subgenomic structures in allo-tetraploid crucian carp hybrids, Hefang crucian carp lines, bluntnose black bream-culter hybrids, gynogenetic largemouth bass, gynogenetic mandarin fish, and their respective parents [257–260].

Combining the established micro-hybrid gynogenetic breeding system with these auxiliary methods, we successfully bred multiple gynogenetic fish lines, including grass carp, blunt snout bream, largemouth bass, mandarin fish, northern snakehead, and marine species such as large yellow croaker. These tools significantly enhanced the success rate and efficiency of gynogenesis.

A patented hatching apparatus for adhesive fish eggs was developed (Figure 6), offering high efficiency, safety, and ease of operation. It effectively mitigates water quality effects, allows



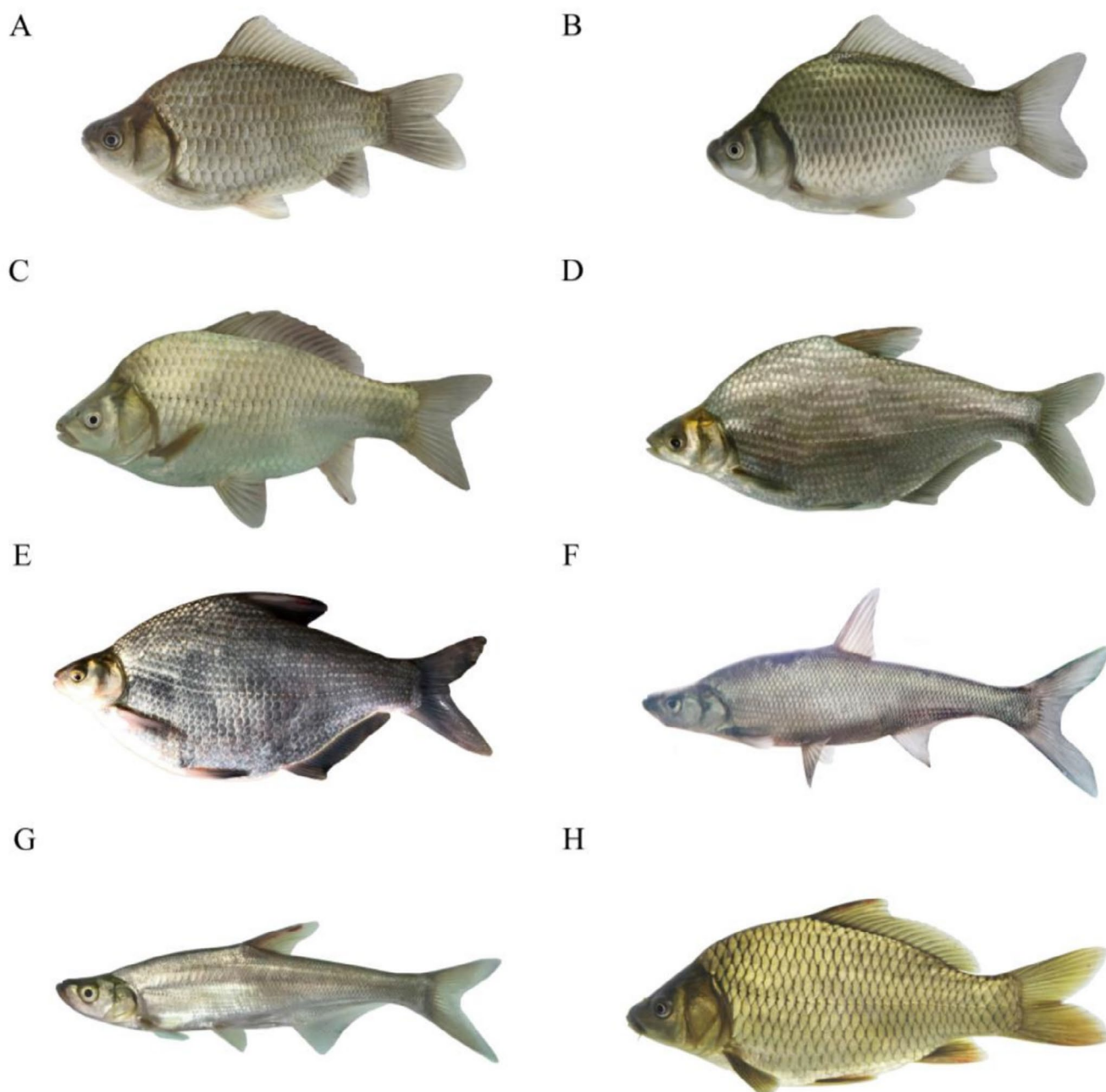
**FIGURE 6** | Drip incubation and temperature-controlled hatching support system.

precise control of incubation temperature, and significantly increases hatch rates. In alignment with ecological principles of “simulating nature and minimizing human intervention,” the team designed an eco-spawning pond that supports natural fertilization, reduces hormone usage, and facilitates the collection and transport of eggs with netting panels [251].

To date, there is no integrated device for gynogenetic (or androgenetic) procedures. Our team has initiated the design and patent process for such equipment, a gynogenesis/androgenesis experimental apparatus. This device offers user-friendly operation, a compact structure, and uniform UV exposure for samples, minimizing external interference and reducing UV harm

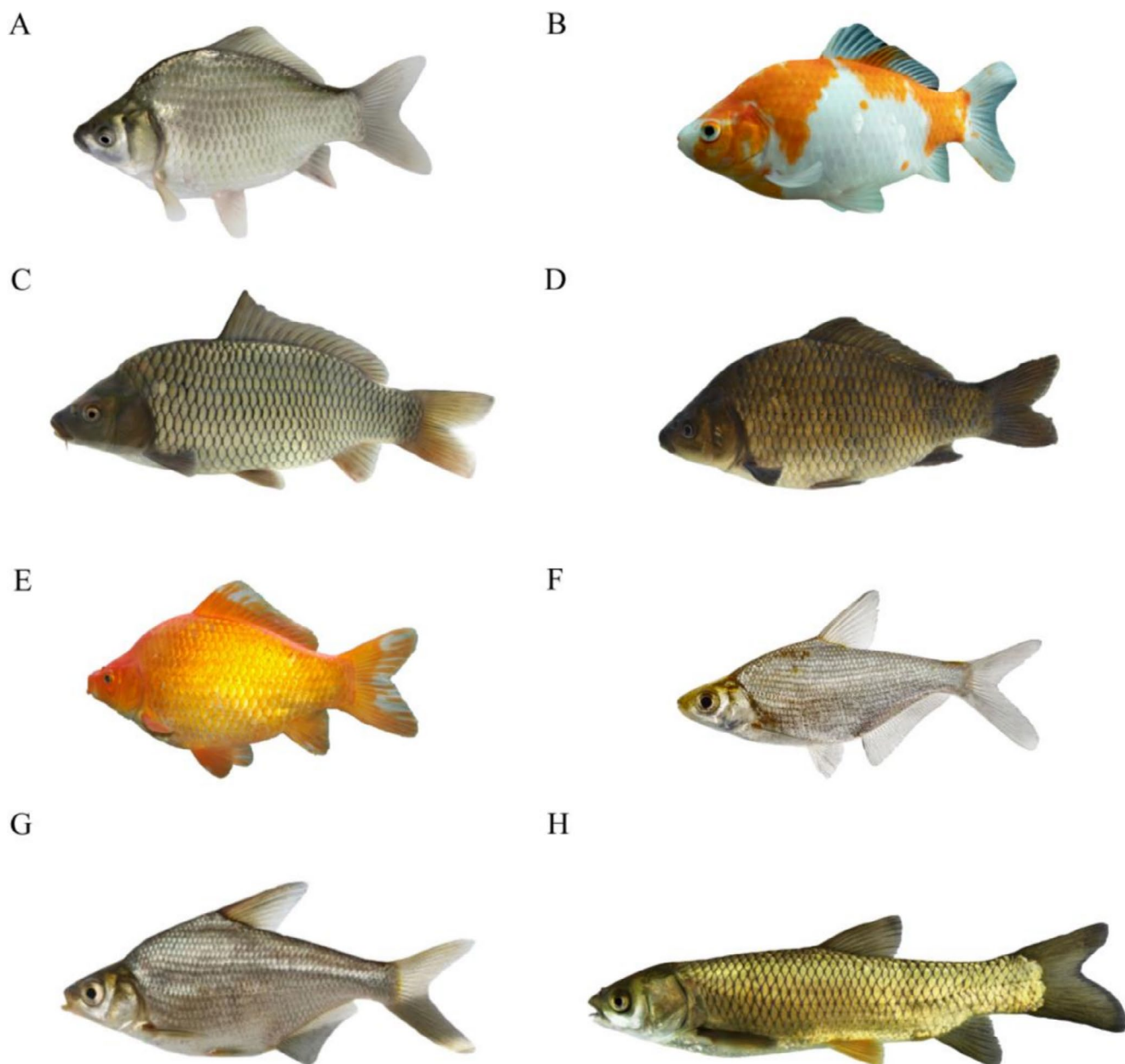
to operators. Its availability will provide convenient and precise support for researchers and breeders.

Through the application of macro-hybridization and micro-hybridization technologies, empowered by these auxiliary breeding techniques, we have successfully developed a series of novel fish strains and premium varieties. This has enriched germplasm resources and enhanced fish population quality. The integrated application of these methods has addressed three key challenges in distant hybridization: (1) low survival, (2) reproductive difficulty, and (3) difficulty in identification. They also solve the two key issues in gynogenesis: (1) low survival and (2) difficulty in population expansion.



**FIGURE 7** | Representative high-quality fish breeds developed via macro-hybrid breeding. (A) Hefang crucian carp; (B) Hefang crucian carp No. 2; (C) Hefang crucian carp No. 3; (D) Hefang Bream; (E) Hefang Bream No. 2; (F) XiangJun culter; (G) XiangJun culter No. 2; (H) Allotriploid carp.





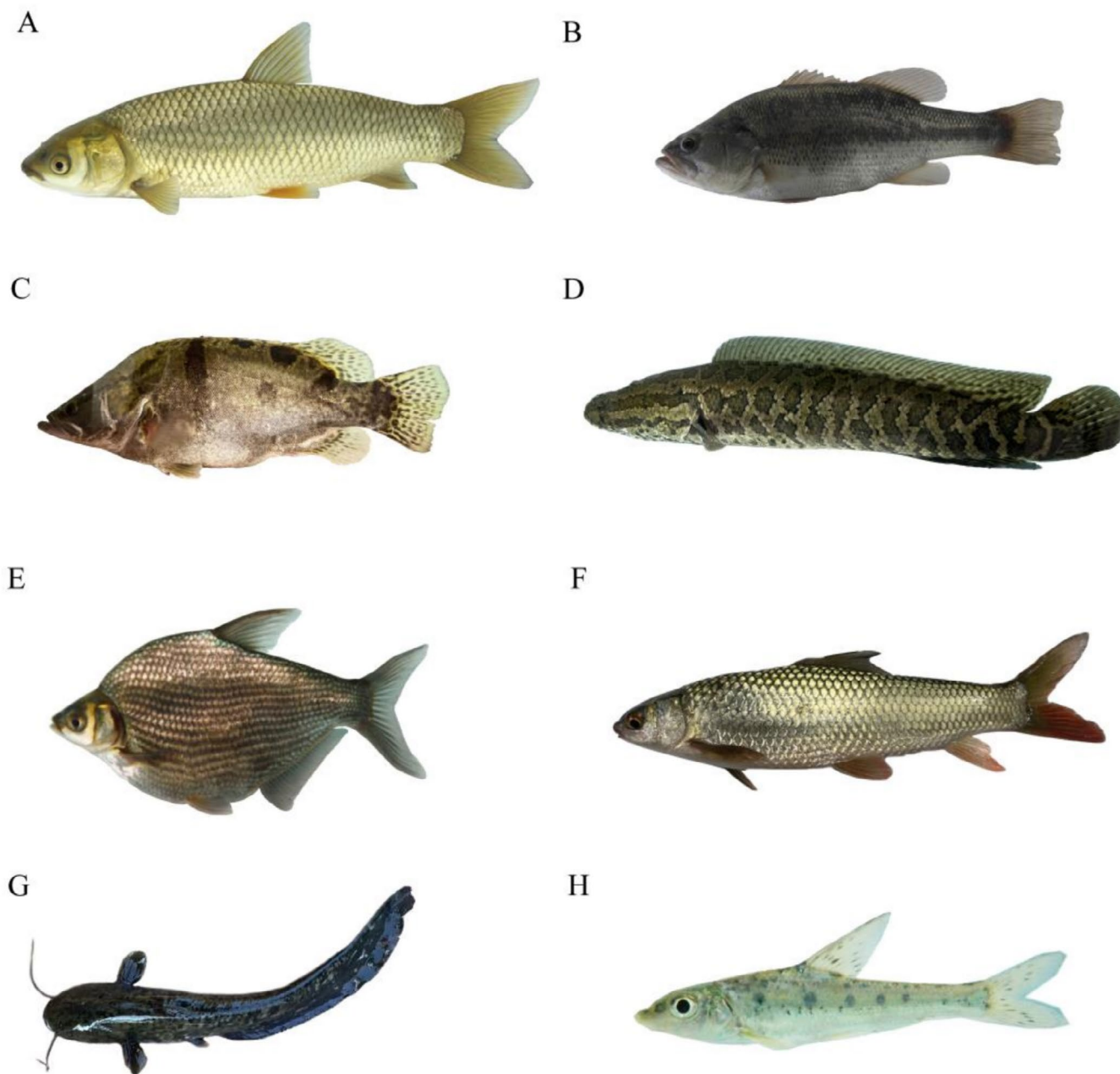
**FIGURE 8** | Representative natural gynogenetic high-quality fish breeds. (A) XiangJun Crucian; (B) XiangJun Flower Crucian; (C) XiangJun Carp; (D) Homotetraploid Crucian (micro-hybrid effect); (E) Red Crucian; (F) Natural gynogenetic bluntnose black bream (bluntnose black bream  $\times$  *Xenocypris davidi* Bleeker); (G) Natural gynogenetic bluntnose black bream (bluntnose black bream  $\times$  mandarin fish); (H) Natural gynogenetic grass carp.

## 7 | Application Cases of Macro-Hybrid and Micro-Hybrid

Over the years, our team has employed macro-hybrid breeding technologies to successfully cultivate a variety of high-quality fish species including Hefang crucian carp (Figure 7A) [261], Hefang crucian carp No. 2 (Figure 7B) [262], Hefang crucian carp No. 3 (Figure 7C) [263], Hefang bream (Figure 7D) [164], Hefang bream No. 2 (Figure 7E) [245], Xiangjun culter (Figure 7F), Xiangjun culter No. 2 (Figure 7G) [253], triploid common carp (Figure 7H) [246]. These diploid and triploid fish exemplify the effectiveness and great potential of macro-hybrid technologies in aquaculture:

**Macro-Hybrid Case: The Hefang crucian carp Series:** This is a typical example of combining one-step and multi-step breeding technologies through the application of macro-hybrid methods. By crossing the Japanese white crucian (female) with the red crucian (male), the  $F_1$  generation was obtained and named Hefang crucian carp. Subsequent self-cross (from  $F_1$  to  $F_2$ ) led to the establishment of a stable line containing both parental subgenomes, classifying it as a macro-hybrid. Hefang crucian carp exhibits a favorable body shape, rapid growth, superior flesh quality, and strong stress resistance [264, 265]. Based on this, Hefang crucian carp was back-crossed with male Japanese white crucian to breed Hefang crucian carp No. 2, which also qualifies as a macro-hybrid, and received national certification





**FIGURE 9** | Representative high-quality fish breeds developed via artificial gynogenesis (micro-hybrid). (A) Disease-resistant gynogenetic grass carp; (B) Gynogenetic largemouth bass; (C) Gynogenetic mandarin fish; (D) Gynogenetic northern snakehead; (E) Gynogenetic bluntnose black bream (koi sperm); (F) Gynogenetic mrigal carp (common carp sperm); (G) Gynogenetic catfish; (H) Gynogenetic *Hemibarbus maculatus*.

in 2022 [266]. Hefang crucian carp No. 2 features a small head, tall back, fast growth, and excellent resilience, making it suitable for various rearing environments (lakes, ponds, paddy fields, lotus fields). With a high content of umami amino acids, it is ideal for soup production. The team developed frozen Hefang crucian carp No. 2 fish soup and commercialized it as a culinary product. A complete supply chain from quality seed to healthy farming and culinary commercialization was established, delivering notable economic, social, and ecological benefits [267]. Hefang crucian carp No. 2 was also crossed with male auto-tetraploid crucian to produce Hefang crucian carp No. 3, a high-quality triploid fish with features including a tall back, barbelless mouth, fast growth, stress resistance, tender flesh, and high amino acid content.

**Macro-Hybrid Case: The Hefang Bream Series:** This represents another successful application of macro-hybrid breeding through the multi-step approach. By hybridizing bluntnose black bream and topmouth culter, the  $F_1$  generation was obtained and then back-crossed with bluntnose black bream to produce a hybrid of topmouth bream. Further back-crossing with female bluntnose black bream yielded Hefang Bream, characterized by a tall back and small head. All these hybrids contained biparental subgenomes, thus qualifying as macro-hybrids. Hefang Bream is well-suited for lakes, ponds, and paddy-lotus fields. Subsequently, a hybrid of topmouth bream was back-crossed with bluntnose black bream to produce Hefang Bream No. 2 (high-back bream), which exhibits rapid growth, low-oxygen tolerance, herbivorous feeding habits,

**TABLE 3** | Natural gynogenetic fish.

Number	Name	Description
1	XiangJun Crucian (Figure 8A)	This novel fish variety, obtained through hybridization between Xiangjiang wild carp and bluntnose black bream, exhibits high survival rates, rapid growth, strong stress resistance, and high nutritional value [161]
2	XiangJun Flower Crucian (Figure 8B)	This stable $F_6$ -generation ornamental crucian hybrid, obtained by crossing ornamental koi carp with bluntnose black bream, demonstrates rapid growth, robust stress tolerance, high fecundity, and superior ornamental quality [153]
3	XiangJun Carp (Figure 8C)	Derived from hybridization between Xiangjiang wild carp and bluntnose black bream, this hybrid variety exhibits good survival rates, rapid growth, a docile temperament, and adaptable feeding behavior, making it well-suited for paddy field culture systems [268]
4	Auto-tetraploid Crucian (Figure 8D)	Developed through hybridization of red crucian carp and bluntnose black bream, this hybrid lineage exhibits stable diploid gamete formation, enhanced stress tolerance, and favorable morphological traits [260]
5	Natural gynogenetic red crucian (Figure 8E)	Originating from hybridization between red crucian carp and bluntnose black bream, this hybrid exhibits rapid growth, superior morphological traits, and stable reproductive performance [260]
6	Natural gynogenetic bluntnose black bream (Figure 8F)	Developed through hybridization of bluntnose black bream and <i>Xenocypris davidi</i> , this hybrid exhibits increased dorsal body depth, high fecundity, and has been stably maintained through self-crossing to the $F_3$ generation [269]
7	Natural gynogenetic bluntnose black bream No. 2 (Figure 8G)	Developed through distant hybridization of bluntnose black bream and mandarin fish, this hybrid exhibits fast growth, high flesh quality, and fertility in both sexes [192]
8	Natural gynogenetic grass carp (Figure 8H)	Developed through distant hybridization of grass carp and topmouth culter, exhibits accelerated somatic growth rates and an enhanced nutritional profile characterized by high protein content and beneficial fatty acid composition [191]

excellent flesh quality, and high protein levels. This strain is ideal for lake and pond culture [164]. Additionally, the hybrid of topmouth bream was back-crossed with topmouth culter to produce XiangJun culter, a macro-hybrid with fast growth, tall body, tender flesh, high meat yield, and strong disease resistance, with 5.7% fewer intermuscular bones than its parent. A further back-cross between XiangJun Blicca and white bream yielded XiangJun culter No. 2, which inherited superior growth and flesh quality traits [253].

In collaboration with Luo Yongju's team from the Guangxi Academy of Fishery Sciences, the author's team also completed the southern breeding of gynogenetic largemouth bass, performing self-cross of back-crossed offspring, laying a solid foundation for large-scale commercial application.

Macro- and micro-hybridization technologies have yielded numerous elite aquatic varieties (Tables 3 and 4). Future advancements require deeper integration with cutting-edge biotechnologies, including the implementation of GS to accelerate trait fixation in hybrid progeny, the application of gene editing to accurately modify disease- and stress-resistance

loci in micro-hybrid lines, and the development of AI-driven predictive models for parental matching. Integrating classical heterosis with molecular design and intelligent prediction tools is crucial for addressing the multifaceted challenges of germplasm innovation efficiency, food safety, and ecological sustainability.

## 8 | Conclusion

Based on our long-term research in fish distant hybridization and gynogenesis, we have for the first time integrated the theories and technologies of these two fields, establishing the conceptual and technical framework of macro-hybrid and micro-hybrid breeding. By synergizing this framework with our previously developed one-step and multi-step breeding strategies, we have successfully generated a series of novel, high-quality fish strains and germplasm resources. This study systematically introduces these innovative theories, technologies, and their practical applications, which we believe will significantly advance the theoretical foundations and technical capabilities of genetic breeding in aquaculture.

**TABLE 4** | Artificial heterologous sperm-induced gynogenetic fish.

Number	Name	Description
1	Gynogenetic grass carp induced by inactivated koi sperm (Figure 9A)	The new variety was back-crossed to wild-type males, resulting in disease-resistant strains characterized by rapid growth, high protein content, low-oxygen tolerance, and herbivorous traits [216, 270]
2	Gynogenetic largemouth bass induced by inactivated mandarin fish sperm (Figure 9B)	The new variety was subjected to iterative back-crossing and self-crossing, resulting in a stable population that preserved the micro-hybrid traits [211]
3	Gynogenetic mandarin fish induced by inactivated largemouth bass sperm (Figure 9C)	The new variety was subjected to successive generations of back-crossing and self-crossing to fix and preserve the favorable traits [208]
4	Gynogenetic northern snakehead induced using inactivated mandarin fish sperm (Figure 9D)	This new variety demonstrated favorable traits, including accelerated growth and improved resilience to abiotic stress [217]
5	Two gynogenetic bluntnose black breamlines induced by inactivated koi and topmouth culter, sperm (Figure 9E)	The new variety demonstrates strong hypoxia tolerance, rapid growth, and enhanced disease resistance [214]
6	Gynogenetic black scraper induced by inactivated common carp sperm (Figure 9F)	The new variety exhibited a 1.4°C reduction in its minimum tolerance temperature, effectively addressing the overwintering challenge in the Hunan region [210]
7	Gynogenetic catfish induced by inactivated spotted channel catfish sperm (Figure 9G)	We observed that the new variety displays micro-hybrid characteristics, accelerated growth, and considerable resilience (data not shown)
8	Gynogenetic <i>Hemibarbus maculatus</i> induced by inactivated koi sperm (Figure 9H)	This new variety is characterized by a combination of advantageous traits, including remarkable tolerance to hypoxic conditions, a high growth rate, desirable flesh characteristics, and robust resistance to common pathogens [209]
9	Gynogenetic white crucian induced by inactivated mirror carp or bluntnose black bream sperm	The new variety is characterized by rapid growth and disease-resistant properties [212]

Looking forward, the breeding system presented here holds substantial potential for broader application. To advance this field, future research should prioritize several key directions: systematic evaluation of macro-hybrid lines for growth performance, flesh quality, and environmental adaptability; enhancement of micro-hybrid breeding efficiency through AI-driven parental matching algorithms integrated with GS; and establishment of comprehensive ecological risk assessment frameworks encompassing long-term monitoring and predictive modeling of genetic introgression. Achieving this vision will demand interdisciplinary collaboration across genetics, bioinformatics, ecology, and policy science, thereby positioning this integrated system as a transformative driver of sustainable innovation in the aquatic seed industry.

#### Author Contributions

**Qizhi Liu:** conceptualization, writing – original draft, writing – review and editing, visualization. **Anmin Liao:** conceptualization, writing – original draft, writing – review and editing, visualization. **Min Tao:** writing – review and editing. **Qinbo Qin:** writing – review and editing. **Kaikun Luo:** writing – review and editing. **Chun Zhang:** writing – review and editing. **Shi Wang:** writing – review and editing. **Yi Zhou:** writing – review and editing. **Fangzhou Hu:** writing – review

and editing. **Yude Wang:** writing – review and editing. **Chang Wu:** writing – review and editing. **Wuhui Li:** writing – review and editing. **Qingfeng Liu:** writing – review and editing. **Chenchen Tang:** writing – review and editing. **Jing Wang:** writing – review and editing. **Rurong Zhao:** writing – review and editing. **Shaojun Liu:** conceptualization, investigation, supervision, funding acquisition, visualization, project administration, resources, writing – original draft, writing – review and editing.

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#### Conflicts of Interest

The authors declare no conflicts of interest.

#### Data Availability Statement

The data that support the findings of this study are available on request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.



## References

1. C. E. Boyd, A. A. McNevin, and R. P. Davis, "The Contribution of Fisheries and Aquaculture to the Global Protein Supply," *Food Security* 14, no. 3 (2022): 1–23, <https://doi.org/10.1007/s12571-021-01246-9>.
2. F. Al Khawli, F. J. Marti-Quijal, E. Ferrer, et al., "Chapter One - Aquaculture and Its By-Products as a Source of Nutrients and Bioactive Compounds," in *Advances in Food and Nutrition Research* (Academic Press, 2020), 1–33.
3. L. L. Wong, S. A. Razali, Z. M. Deris, et al., "Application of Second-Generation Sequencing (SGS) and Third Generation Sequencing (TGS) in Aquaculture Breeding Program," *Aquaculture* 548 (2022): 737633, <https://doi.org/10.1016/j.aquaculture.2021.737633>.
4. A. A. Gheyas, J. A. Woolliams, J. B. Taggart, et al., "Heritability Estimation of Silver Carp (*Hypophthalmichthys molitrix*) Harvest Traits Using Microsatellite Based Parentage Assignment," *Aquaculture* 294, no. 3 (2009): 187–193, <https://doi.org/10.1016/j.aquaculture.2009.06.013>.
5. M. A. Rezk, R. O. Smitherman, J. C. Williams, A. Nichols, H. Kucuktas, and R. A. Dunham, "Response to Three Generations of Selection for Increased Body Weight in Channel Catfish, *Ictalurus punctatus*, Grown in Earthen Ponds," *Aquaculture* 228, no. 1 (2003): 69–79, [https://doi.org/10.1016/S0044-8486\(03\)00216-3](https://doi.org/10.1016/S0044-8486(03)00216-3).
6. G. C. Embury and C. O. Hayford, "The Advantage of Rearing Brook Trout Fingerlings From Selected Breeders," *Transactions of the American Fisheries Society* 55, no. 1 (1925): 135–148, [https://doi.org/10.1577/1548-8659\(1925\)55\[135:TAORBT\]2.0.CO;2](https://doi.org/10.1577/1548-8659(1925)55[135:TAORBT]2.0.CO;2).
7. E. Saillant, M. Dupont-Nivet, P. Haffray, and B. Chatain, "Estimates of Heritability and Genotype–Environment Interactions for Body Weight in Sea Bass (*Dicentrarchus labrax* L.) Raised Under Communal Rearing Conditions," *Aquaculture* 254, no. 1 (2006): 139–147, <https://doi.org/10.1016/j.aquaculture.2005.10.018>.
8. A. Kaase, O. Ritola, T. Paananen, H. Wahlroos, and E. A. Mäntysaari, "Genetic Trends in Growth, Sexual Maturity and Skeletal Deformations, and Rate of Inbreeding in a Breeding Programme for Rainbow Trout (*Oncorhynchus mykiss*)," *Aquaculture* 247, no. 1 (2005): 177–187, <https://doi.org/10.1016/j.aquaculture.2005.02.023>.
9. R. Neira, N. F. Díaz, G. A. E. Gall, J. A. Gallardo, J. P. Lhorente, and A. Alert, "Genetic Improvement in Coho Salmon (*Oncorhynchus kisutch*). II: Selection Response for Early Spawning Date," *Aquaculture* 257, no. 1 (2006): 1–8, <https://doi.org/10.1016/j.aquaculture.2006.03.001>.
10. J. Abwao, J. Jung'a, J. E. Barasa, et al., "Selective Breeding of Nile Tilapia, *Oreochromis niloticus*: A Strategy for Increased Genetic Diversity and Sustainable Development of Aquaculture in Kenya," *Journal of Applied Aquaculture* 35, no. 2 (2023): 237–256, <https://doi.org/10.1080/10454438.2021.1958728>.
11. Z. Dong, N. H. Nguyen, and W. Zhu, "Genetic Evaluation of a Selective Breeding Program for Common Carp *Cyprinus carpio* Conducted From 2004 to 2014," *BMC Genetics* 16, no. 1 (2015): 94, <https://doi.org/10.1186/s12863-015-0256-2>.
12. B.-H. Nam, D. Yoo, Y.-O. Kim, et al., "Whole Genome Sequencing Reveals the Impact of Recent Artificial Selection on Red Sea Bream Reared in Fish Farms," *Scientific Reports* 9, no. 1 (2019): 6487, <https://doi.org/10.1038/s41598-019-42988-z>.
13. Y. Ning, X. Liu, Z. Y. Wang, W. Guo, Y. Li, and F. Xie, "A Genetic Map of Large Yellow Croaker *Pseudosciaena crocea*," *Aquaculture* 264, no. 1 (2007): 16–26, <https://doi.org/10.1016/j.aquaculture.2006.12.042>.
14. Y.-G. Liu, S.-L. Chen, B.-F. Li, Z.-J. Wang, and Z. Liu, "Analysis of Genetic Variation in Selected Stocks of Hatchery Flounder, *Paralichthys olivaceus*, Using AFLP Markers," *Biochemical Systematics and Ecology* 33, no. 10 (2005): 993–1005, <https://doi.org/10.1016/j.bse.2005.02.008>.
15. T. Zhang, J. Kong, B. Liu, et al., "Genetic Parameter Estimation for Juvenile Growth and Upper Thermal Tolerance in Turbot (*Scophthalmus maximus* Linnaeus)," *Acta Oceanologica Sinica* 33, no. 8 (2014): 106–110, <https://doi.org/10.1007/s13131-014-0460-3>.
16. W. Hong and Q. Zhang, "Review of Captive Bred Species and Fry Production of Marine Fish in China," *Aquaculture* 227, no. 1 (2003): 305–318, [https://doi.org/10.1016/S0044-8486\(03\)00511-8](https://doi.org/10.1016/S0044-8486(03)00511-8).
17. M. Agarwal, N. Shrivastava, and H. Padh, "Advances in Molecular Marker Techniques and Their Applications in Plant Sciences," *Plant Cell Reports* 27, no. 4 (2008): 617–631, <https://doi.org/10.1007/s00299-008-0507-z>.
18. W. Yang, X. Kang, Q. Yang, Y. Lin, and M. Fang, "Review on the Development of Genotyping Methods for Assessing Farm Animal Diversity," *Journal of Animal Science and Biotechnology* 4, no. 1 (2013): 2, <https://doi.org/10.1186/2049-1891-4-2>.
19. J. H. Postlethwait, S. L. Johnson, C. N. Midson, et al., "A Genetic Linkage Map for the Zebrafish," *Science* 264, no. 5159 (1994): 699–703, <https://doi.org/10.1126/science.8171321>.
20. K. Naruse, M. Tanaka, K. Mita, A. Shima, J. Postlethwait, and H. Mitani, "A Medaka Gene Map: The Trace of Ancestral Vertebrate Proto-Chromosomes Revealed by Comparative Gene Mapping," *Genome Research* 14, no. 5 (2004): 820–828, <https://doi.org/10.1101/gr.2004004>.
21. W. Kai, K. Kikuchi, M. Fujita, et al., "A Genetic Linkage Map for the Tiger Pufferfish, *Takifugu rubripes*," *Genetics* 171, no. 1 (2005): 227–238, <https://doi.org/10.1534/genetics.105.042051>.
22. R. B. Walter, J. D. Rains, J. E. Russell, et al., "A Microsatellite Genetic Linkage Map for Xiphophorus," *Genetics* 168, no. 1 (2004): 363–372, <https://doi.org/10.1534/genetics.103.019349>.
23. C. L. Peichel, K. S. Nereng, K. A. Ohgi, et al., "The Genetic Architecture of Divergence Between Threespine Stickleback Species," *Nature* 414, no. 6866 (2001): 901–905, <https://doi.org/10.1038/414901a>.
24. W. P. Young, P. A. Wheeler, V. H. Coryell, P. Keim, and G. H. Thorgaard, "A Detailed Linkage Map of Rainbow Trout Produced Using Doubled Haploids," *Genetics* 148, no. 2 (1998): 839–850, <https://doi.org/10.1093/genetics/148.2.839>.
25. B. Y. Lee, W. J. Lee, J. T. Streelman, et al., "A Second-Generation Genetic Linkage Map of Tilapia (*Oreochromis* spp.)," *Genetics* 170, no. 1 (2005): 237–244, <https://doi.org/10.1534/genetics.104.035022>.
26. V. L. Ramya and B. K. Behera, "Molecular Markers and Their Application in Fisheries and Aquaculture," in *Biotechnological Tools in Fisheries and Aquatic Health Management* (Springer Nature Singapore, 2023), 115–150.
27. H. Liu, B. Guan, J. Xu, C. Hou, H. Tian, and H. Chen, "Genetic Manipulation of Sex Ratio for the Large-Scale Breeding of YY Super-Male and XY All-Male Yellow Catfish (*Pelteobagrus fulvidraco* (Richardson)), " *Marine Biotechnology (New York, N.Y.)* 15, no. 3 (2013): 321–328, <https://doi.org/10.1007/s10126-012-9487-7>.
28. Z. J. Liu and J. F. Cordes, "DNA Marker Technologies and Their Applications in Aquaculture Genetics," *Aquaculture* 238, no. 1 (2004): 1–37, <https://doi.org/10.1016/j.aquaculture.2004.05.027>.
29. M. Y. Laghari, P. Lashari, X. Zhang, et al., "QTL Mapping for Economically Important Traits of Common Carp (*Cyprinus carpio* L.)," *Journal of Applied Genetics* 56, no. 1 (2015): 65–75, <https://doi.org/10.1007/s13353-014-0232-y>.
30. M. Ou, K.-C. Chen, Q. Luo, et al., "Performance Evaluation of XY All-Male Hybrids Derived From XX Female Channa Argus and YY Super-Males Channa Maculate," *Aquaculture Reports* 20 (2021): 100768, <https://doi.org/10.1016/j.aqrep.2021.100768>.
31. D. Wang, H.-L. Mao, H.-X. Chen, H.-Q. Liu, and J.-F. Gui, "Isolation of Y- and X-Linked SCAR Markers in Yellow Catfish and Application in the Production of All-Male Populations," *Animal Genetics* 40, no. 6 (2009): 978–981, <https://doi.org/10.1111/j.1365-2052.2009.01941.x>.



32. C. Dan, J. Mei, D. Wang, and J. F. Gui, "Genetic Differentiation and Efficient Sex-Specific Marker Development of a Pair of Y- and X-Linked Markers in Yellow Catfish," *International Journal of Biological Sciences* 9, no. 10 (2013): 1043–1049, <https://doi.org/10.7150/ijbs.7203>.
33. Q. Yu, S. Liu, Q. Zhu, R. Chen, W. Hu, and D. Xu, "Genetic Variability of Mass-Selected and Wild Populations of Yellow Drum (*Nibea albiglora*) Revealed Using Microsatellites," *Fishes* 9 (2024): 25, <https://doi.org/10.3390/fishes9010025>.
34. J. H. Xia, Z. Bai, Z. Meng, et al., "Signatures of Selection in Tilapia Revealed by Whole Genome Resequencing," *Scientific Reports* 5 (2015): 14168, <https://doi.org/10.1038/srep14168>.
35. X. Zhao, T. Zheng, T. Gao, and N. Song, "Whole-Genome Resequencing Reveals Genetic Diversity and Selection Signals in Warm Temperate and Subtropical Sillago Sinica Populations," *BMC Genomics* 24, no. 1 (2023): 547, <https://doi.org/10.1186/s12864-023-09652-3>.
36. E. Sarropoulou, D. Noudili, A. Magoulas, and G. Kotoulas, "Linking the Genomes of Nonmodel Teleosts Through Comparative Genomics," *Marine Biotechnology* 10, no. 3 (2008): 227–233, <https://doi.org/10.1007/s10126-007-9066-5>.
37. B. Star, A. J. Nederbragt, S. Jentoft, et al., "The Genome Sequence of Atlantic Cod Reveals a Unique Immune System," *Nature* 477, no. 7363 (2011): 207–210, <https://doi.org/10.1038/nature10342>.
38. S. Chen, G. Zhang, C. Shao, et al., "Whole-Genome Sequence of a Flatfish Provides Insights Into ZW Sex Chromosome Evolution and Adaptation to a Benthic Lifestyle," *Nature Genetics* 46, no. 3 (2014): 253–260, <https://doi.org/10.1038/ng.2890>.
39. G. Zhang, X. Fang, X. Guo, et al., "The Oyster Genome Reveals Stress Adaptation and Complexity of Shell Formation," *Nature* 490, no. 7418 (2012): 49–54, <https://doi.org/10.1038/nature11413>.
40. P. Xu, X. Zhang, X. Wang, et al., "Genome Sequence and Genetic Diversity of the Common Carp, *Cyprinus carpio*," *Nature Genetics* 46, no. 11 (2014): 1212–1219, <https://doi.org/10.1038/ng.3098>.
41. A. A. Gheyas, Y. Basavaraju, B. J. McAndrew, and D. J. Penman, "Monitoring Genetic Diversity in a Mass Selection Programme of Common Carp (*Cyprinus carpio*) Using Microsatellite Markers," *Aquaculture* 272, no. S1 (2007): S261–S262, <https://doi.org/10.1016/j.aquaculture.2007.07.072>.
42. J. D'Ambrosio, R. Morvezen, S. Brard-Fudulea, et al., "Genetic Architecture and Genomic Selection of Female Reproduction Traits in Rainbow Trout," *BMC Genomics* 21, no. 1 (2020): 558, <https://doi.org/10.1186/s12864-020-06955-7>.
43. N. Li, T. Zhou, X. Geng, et al., "Identification of Novel Genes Significantly Affecting Growth in Catfish Through GWAS Analysis," *Molecular Genetics and Genomics* 293, no. 3 (2018): 587–599, <https://doi.org/10.1007/s00438-017-1406-1>.
44. X. Geng, S. Liu, Z. Yuan, Y. Jiang, D. Zhi, and Z. Liu, "A Genome-Wide Association Study Reveals That Genes With Functions for Bone Development Are Associated With Body Conformation in Catfish," *Marine Biotechnology* 19, no. 6 (2017): 570–578, <https://doi.org/10.1007/s10126-017-9775-3>.
45. Z. Gao, C. Zheng, Y. Zhang, et al., "Genome-Wide Analysis of DNA Methylation and Gene Expression in Large Yellow Croaker (*Larimichthys crocea*) Under Hypoxic Stress," *Aquaculture* 595 (2025): 741624, <https://doi.org/10.1016/j.aquaculture.2024.741624>.
46. B. L. Langille, P. Sae-Lim, S. Boisson, P. G. Wiper, and A. F. Garber, "Genome-Wide Association Identifies Genomic Regions Influencing Fillet Color in Northwest Atlantic Salmon (*Salmo salar* Linnaeus 1758)," *Frontiers in Genetics* 15 (2024): 1402927, <https://doi.org/10.3389/fgene.2024.1402927>.
47. C. Yu, Y. Jiang, C. Zhang, et al., "Whole-Genome Resequencing of Grass Carp (*Ctenopharyngodon idella*) for Genome-Wide Association Study on GCRV Resistance," *Aquaculture* 592 (2024): 741243, <https://doi.org/10.1016/j.aquaculture.2024.741243>.
48. S. Xu, "Introduction to Quantitative Genetics," in *Quantitative Genetics* (Springer International Publishing, 2022), 1–12.
49. W. G. Hill, "Applications of Population Genetics to Animal Breeding, From Wright, Fisher and Lush to Genomic Prediction," *Genetics* 196, no. 1 (2014): 1–16, <https://doi.org/10.1534/genetics.112.147850>.
50. R. Lande and R. Thompson, "Efficiency of Marker-Assisted Selection in the Improvement of Quantitative Traits," *Genetics* 124, no. 3 (1990): 743–756, <https://doi.org/10.1093/genetics/124.3.743>.
51. D. o. Agronomy and P. G. U. o. M. S. P. MN, "Molecular Markers and Selection for Complex Traits in Plants: Learning From the Last 20 Years," *Crop Science* 48, no. 5 (2008): 1649–1664, <https://doi.org/10.2135/cropsci2008.03.0131>.
52. J. C. M. Dekkers, "Commercial Application of Marker- and Gene-Assisted Selection in Livestock: Strategies and Lessons," *Journal of Animal Science* 82, no. 13 (2004): E313–E328, [https://doi.org/10.2527/2004.8213\\_supplE313x](https://doi.org/10.2527/2004.8213_supplE313x).
53. T. H. Meuwissen, B. J. Hayes, and M. E. Goddard, "Prediction of Total Genetic Value Using Genome-Wide Dense Marker Maps," *Genetics* 157, no. 4 (2001): 1819–1829, <https://doi.org/10.1093/genetics/157.4.1819>.
54. B. J. Hayes, P. J. Bowman, A. J. Chamberlain, and M. E. Goddard, "Invited Review: Genomic Selection in Dairy Cattle: Progress and Challenges," *Journal of Dairy Science* 92, no. 2 (2009): 433–443, <https://doi.org/10.3168/jds.2008-1646>.
55. S. Y. Yan, D. Y. Lu, M. Du, et al., "Nuclear Transplantation in Teleosts. Hybrid Fish From the Nucleus of Crucian and the Cytoplasm of Carp," *Scientia Sinica. Series B* 27, no. 10 (1984): 1029–1034.
56. S. Y. Yan, M. Tu, H. Y. Yang, et al., "Developmental Incompatibility Between Cell Nucleus and Cytoplasm as Revealed by Nuclear Transplantation Experiments in Teleost of Different Families and Orders," *International Journal of Developmental Biology* 34, no. 2 (1990): 255–266.
57. Y. Takeuchi, G. Yoshizaki, and T. Takeuchi, "Generation of Live Fry From Intraperitoneally Transplanted Primordial Germ Cells in Rainbow Trout," *Biology of Reproduction* 69, no. 4 (2003): 1142–1149, <https://doi.org/10.1095/biolreprod.103.017624>.
58. Y. Takeuchi, G. Yoshizaki, and T. Takeuchi, "Surrogate Broodstock Produces Salmonids," *Nature* 430, no. 7000 (2004): 629–630, <https://doi.org/10.1038/430629a>.
59. Y. Hong, T. Liu, H. Zhao, et al., "Establishment of a Normal Medakafish Spermatogonial Cell Line Capable of Sperm Production In Vitro," *Proceedings of the National Academy of Sciences* 101, no. 21 (2004): 8011–8016, <https://doi.org/10.1073/pnas.0308668101>.
60. T. Okutsu, K. Suzuki, Y. Takeuchi, T. Takeuchi, and G. Yoshizaki, "Testicular Germ Cells Can Colonize Sexually Undifferentiated Embryonic Gonad and Produce Functional Eggs in Fish," *Proceedings of the National Academy of Sciences of the United States of America* 103, no. 8 (2006): 2725–2729, <https://doi.org/10.1073/pnas.0509218103>.
61. M. Yi, N. Hong, and Y. Hong, "Generation of Medaka Fish Haploid Embryonic Stem Cells," *Science* 326, no. 5951 (2009): 430–433, <https://doi.org/10.1126/science.1175151>.
62. R. H. Nóbrega, C. D. Greebe, H. van de Kant, J. Bogerd, L. R. de França, and R. W. Schulz, "Spermatogonial Stem Cell Niche and Spermatogonial Stem Cell Transplantation in Zebrafish," *PLoS One* 5, no. 9 (2010): 12808, <https://doi.org/10.1371/journal.pone.0012808>.
63. S. K. Majhi, R. S. Hattori, M. Yokota, S. Watanabe, and C. A. Strüssmann, "Germ Cell Transplantation Using Sexually Competent Fish: An Approach for Rapid Propagation of Endangered and Valuable

- Germlines," *PLoS One* 4, no. 7 (2009): e6132, <https://doi.org/10.1371/journal.pone.0006132>.
64. T. Okutsu, S. Shikina, M. Kanno, Y. Takeuchi, and G. Yoshizaki, "Production of Trout Offspring From Triploid Salmon Parents," *Science* 317, no. 5844 (2007): 1517, <https://doi.org/10.1126/science.1145626>.
65. X. G. Wang, Z. Y. Zhu, Y. H. Sun, and J. Zhao, "Nuclear Transfer and Reprogramming in Fish," *Yi Chuan* 35, no. 4 (2013): 433–440, <https://doi.org/10.3724/sp.j.1005.2013.00433>.
66. H. Chen, Y. Yi, M. Chen, and X. Yang, "Studies on the Developmental Potentiality of Cultured Cell Nuclei of Fish," *International Journal of Biological Sciences* 6, no. 2 (2010): 192–198, <https://doi.org/10.7150/ijbs.6.192>.
67. Z. Y. Zhu and Y. H. Sun, "Embryonic and Genetic Manipulation in Fish," *Cell Research* 10, no. 1 (2000): 17–27, <https://doi.org/10.1038/sj.cr.7290032>.
68. Y. Sun, S. Chen, Y. Wang, and Z. Zhu, "The Onset of Foreign Gene Transcription in Nuclear-Transferred Embryos of Fish," *Science in China Series C: Life Sciences* 43, no. 6 (2000): 597–605, <https://doi.org/10.1007/BF02882280>.
69. H. Ye, C.-J. Li, H.-M. Yue, et al., "Establishment of Intraperitoneal Germ Cell Transplantation for Critically Endangered Chinese Sturgeon *Acipenser sinensis*," *Theriogenology* 94 (2017): 37–47, <https://doi.org/10.1016/j.theriogenology.2017.02.009>.
70. H. Ye, Y. Takeuchi, H. Du, et al., "Spermatogonia From Cryopreserved Testes of Critically Endangered Chinese Sturgeon Efficiently Colonized and Preferentially Proliferated in the Recipient Gonads of Yangtze Sturgeon," *Marine Biotechnology* 24, no. 1 (2022): 136–150, <https://doi.org/10.1007/s10126-022-10092-5>.
71. M. C. Mullins, "A Toast to a Zebrafish Germ Cell Induction Cocktail," *Science China Life Sciences* 67, no. 6 (2024): 1314–1315, <https://doi.org/10.1007/s11427-024-2530-6>.
72. T. Saito, R. Goto-Kazeto, K. Arai, and E. Yamaha, "Xenogenesis in Teleost Fish Through Generation of Germ-Line Chimeras by Single Primordial Germ Cell Transplantation," *Biology of Reproduction* 78, no. 1 (2008): 159–166, <https://doi.org/10.1095/biolreprod.107.060038>.
73. S. Lacerda, S. Batlouni, S. Silva, C. Homem, and L. França, "Germ Cells Transplantation in Fish: The Nile-Tilapia Model," *Animal Reproduction* 2 (2006): 146–159.
74. X. Wang, J. Zhu, H. Wang, et al., "Induced Formation of Primordial Germ Cells From Zebrafish Blastomeres by Germplasm Factors," *Nature Communications* 14, no. 1 (2023): 7918, <https://doi.org/10.1038/s41467-023-43587-3>.
75. Z. Chen, M. He, H. Wang, et al., "Intestinal DHA-PA-PG Axis Promotes Digestive Organ Expansion by Mediating Usage of Maternally Deposited Yolk Lipids," *Nature Communications* 15, no. 1 (2024): 9769, <https://doi.org/10.1038/s41467-024-54258-2>.
76. F. Zhang, Y. Hao, X. Li, et al., "Surrogate Production of Genome-Edited Sperm From a Different Subfamily by Spermatogonial Stem Cell Transplantation," *Science China Life Sciences* 65, no. 5 (2022): 969–987, <https://doi.org/10.1007/s11427-021-1989-9>.
77. D. Ye, L. Zhu, Q. Zhang, et al., "Abundance of Early Embryonic Primordial Germ Cells Promotes Zebrafish Female Differentiation as Revealed by Lifetime Labeling of Germline," *Marine Biotechnology* 21, no. 2 (2019): 217–228, <https://doi.org/10.1007/s10126-019-09874-1>.
78. R. Yazawa, Y. Takeuchi, K. Higuchi, T. Yatabe, N. Kabeya, and G. Yoshizaki, "Chub Mackerel Gonads Support Colonization, Survival, and Proliferation of Intraperitoneally Transplanted Xenogenic Germ Cells," *Biology of Reproduction* 82, no. 5 (2010): 896–904, <https://doi.org/10.1095/biolreprod.109.081281>.
79. D. J. Luo, W. Hu, S. P. Chen, and Z. Y. Zhu, "Critical Developmental Stages for the Efficiency of Somatic Cell Nuclear Transfer in Zebrafish," *International Journal of Biological Sciences* 7, no. 4 (2011): 476–486, <https://doi.org/10.7150/ijbs.7.476>.
80. H. Ye, C. Zhou, H. Yue, et al., "Cryopreservation of Germline Stem Cells in American Paddlefish (*Polyodon spathula*)," *Animal Reproduction Science* 224 (2021): 106667, <https://doi.org/10.1016/j.anireprosci.2020.106667>.
81. Y. Wakamatsu, B. Ju, I. Pristayaznhyuk, et al., "Fertile and Diploid Nuclear Transplants Derived From Embryonic Cells of a Small Laboratory Fish, Medaka (*Oryzias latipes*)," *Proceedings of the National Academy of Sciences of the United States of America* 98, no. 3 (2001): 1071–1076, <https://doi.org/10.1073/pnas.98.3.1071>.
82. L. N. West-Livingston, J. Park, S. J. Lee, A. Atala, and J. J. Yoo, "The Role of the Microenvironment in Controlling the Fate of Bioprinted Stem Cells," *Chemical Reviews* 120, no. 19 (2020): 11056–11092, <https://doi.org/10.1021/acs.chemrev.0c00126>.
83. Y. H. Jin, D. Robledo, J. M. Hickey, M. J. McGrew, and R. D. Houston, "Surrogate Broodstock to Enhance Biotechnology Research and Applications in Aquaculture," *Biotechnology Advances* 49 (2021): 107756, <https://doi.org/10.1016/j.biotechadv.2021.107756>.
84. D. Chourrout, "Tetraploidy Induced by Heat Shocks in the Rainbow Trout (*Salmo Gairdneri* R.)," *Reproduction Nutrition Development* 22, no. 3 (1982): 569–574, <https://doi.org/10.1051/rnd:19820412>.
85. C. A. Bidwell, C. Larry Chrisman, and G. S. Libey, "Polyploidy Induced by Heat Shock in Channel Catfish," *Aquaculture* 51, no. 1 (1985): 25–32, [https://doi.org/10.1016/0044-8486\(85\)90237-6](https://doi.org/10.1016/0044-8486(85)90237-6).
86. M. Flajšhans, O. Linhart, and P. Kvasnička, "Genetic Studies of Tench (*Tinca tinca* L.): Induced Triploidy and Tetraploidy and First Performance Data," *Aquaculture* 113, no. 4 (1993): 301–312, [https://doi.org/10.1016/0044-8486\(93\)90401-J](https://doi.org/10.1016/0044-8486(93)90401-J).
87. Y. K. Nam, G. C. Choi, D. J. Park, and D. S. Kim, "Survival and Growth of Induced Tetraploid Mud Loach," *Aquaculture International* 9, no. 1 (2001): 61–71, <https://doi.org/10.1023/A:1012540024333>.
88. J. M. Myers, "Tetraploid Induction in *Oreochromis* spp," *Aquaculture* 57, no. 1 (1986): 281–287, [https://doi.org/10.1016/0044-8486\(86\)90206-1](https://doi.org/10.1016/0044-8486(86)90206-1).
89. T. Hayashida, K. Higuchi, K. Nomura, et al., "Optimization of Cold-Shock Conditions for the Induction of Triploidy in the Pacific Bluefin Tuna, *Thunnus orientalis* (Temminck et Schlegel)," *Aquaculture* 530 (2021): 735769, <https://doi.org/10.1016/j.aquaculture.2020.735769>.
90. A. Hassan, V. T. Okomoda, and P. J. Pradeep, "Triploidy Induction by Electric Shock in Red Hybrid Tilapia," *Aquaculture* 495 (2018): 823–830, <https://doi.org/10.1016/j.aquaculture.2018.06.074>.
91. M. Tao, Y. Zhou, S. Li, et al., "MicroRNA Alternations in the Testes Related to the Sterility of Triploid Fish," *Marine Biotechnology* 20, no. 6 (2018): 739–749, <https://doi.org/10.1007/s10126-018-9845-1>.
92. T. A. Delomas and K. Dabrowski, "Why Are Triploid Zebrafish All Male?," *Molecular Reproduction and Development* 85, no. 7 (2018): 612–621, <https://doi.org/10.1002/mrd.22998>.
93. L. Zhou and J. Gui, "Natural and Artificial Polyploids in Aquaculture," *Aquaculture and Fisheries* 2, no. 3 (2017): 103–111, <https://doi.org/10.1016/j.aaf.2017.04.003>.
94. K. Siripattarapavat, S. Prukudom, and J. Cibelli, "Method for Somatic Cell Nuclear Transfer in Zebrafish," *Methods in Cell Biology* 135 (2016): 245–257, <https://doi.org/10.1016/bs.mcmb.2016.04.022>.
95. R. Yazawa, Y. Takeuchi, Y. Machida, et al., "Production of Triploid Eastern Little Tuna, *Euthynnus affinis* (Cantor, 1849)," *Aquaculture Research* 50, no. 5 (2019): 1422–1430, <https://doi.org/10.1111/are.14017>.
96. R. S. Rasmussen and M. T. Morrissey, "Biotechnology in Aquaculture: Transgenics and Polyploidy," *Comprehensive Reviews in Food Science and Food Safety* 6, no. 1 (2007): 2–16, <https://doi.org/10.1111/j.1541-4337.2007.00013.x>.

97. Z. Zhu, L. He, and S. Chen, "Novel Gene Transfer Into the Fertilized Eggs of Gold Fish (*Carassius auratus* L. 1758)," *Journal of Applied Ichthyology* 1, no. 1 (1985): 31–34, <https://doi.org/10.1111/j.1439-0426.1985.tb00408.x>.
98. F. Yu, J. Xiao, X. Liang, et al., "Rapid Growth and Sterility of Growth Hormone Gene Transgenic Triploid Carp," *Chinese Science Bulletin* 56, no. 16 (2011): 1679–1684, <https://doi.org/10.1007/s11434-011-4446-7>.
99. Z. Zhu, L. He, and T. T. Chen, "Primary-Structural and Evolutionary Analyses of the Growth-Hormone Gene From Grass Carp (*Ctenopharyngodon idellus*)," *European Journal of Biochemistry* 207, no. 2 (1992): 643–648, <https://doi.org/10.1111/j.1432-1033.1992.tb17091.x>.
100. Y. H. Sun, S. P. Chen, Y. P. Wang, W. Hu, and Z. Y. Zhu, "Cytoplasmic Impact on Cross-Genus Cloned Fish Derived From Transgenic Common Carp (*Cyprinus carpio*) Nuclei and Goldfish (*Carassius auratus*) Enucleated Eggs," *Biology of Reproduction* 72, no. 3 (2005): 510–515, <https://doi.org/10.1095/biolreprod.104.031302>.
101. D. Li, C. Fu, W. Hu, S. Zhong, Y. Wang, and Z. Zhu, "Rapid Growth Cost in 'All-Fish' Growth Hormone Gene Transgenic Carp: Reduced Critical Swimming Speed," *Chinese Science Bulletin* 52, no. 11 (2007): 1501–1506, <https://doi.org/10.1007/s11434-007-0217-x>.
102. M. Duan, T. Zhang, W. Hu, et al., "Behavioral Alterations in GH Transgenic Common Carp May Explain Enhanced Competitive Feeding Ability," *Aquaculture* 317, no. 1 (2011): 175–181, <https://doi.org/10.1016/j.aquaculture.2011.04.013>.
103. L. M. Houdebine and D. Chourrout, "Transgenesis in Fish," *Experientia* 47, no. 9 (1991): 891–897, <https://doi.org/10.1007/bf01929879>.
104. M. Rembold, K. Lahiri, N. S. Foulkes, and J. Wittbrodt, "Transgenesis in Fish: Efficient Selection of Transgenic Fish by Co-Injection With a Fluorescent Reporter Construct," *Nature Protocols* 1, no. 3 (2006): 1133–1139, <https://doi.org/10.1038/nprot.2006.165>.
105. C. Fu, Y. Cui, S. S. O. Hung, and Z. Zhu, "Growth and Feed Utilization by F4 Human Growth Hormone Transgenic Carp Fed Diets With Different Protein Levels," *Journal of Fish Biology* 53, no. 1 (1998): 115–129, <https://doi.org/10.1111/j.1095-8649.1998.tb00114.x>.
106. T. Zhu, T. Zhang, Y. Wang, Y. Chen, W. Hu, and Z. Zhu, "Effects of Growth Hormone (GH) Transgene and Nutrition on Growth and Bone Development in Common Carp," *Journal of Experimental Zoology. Part A, Ecological Genetics and Physiology* 319, no. 8 (2013): 451–460, <https://doi.org/10.1002/jez.1808>.
107. Y. K. Nam, J. K. Noh, Y. S. Cho, et al., "Dramatically Accelerated Growth and Extraordinary Gigantism of Transgenic Mud Loach *Misgurnus mizolepis*," *Transgenic Research* 10, no. 4 (2001): 353–362, <https://doi.org/10.1023/A:1016696104185>.
108. M. Cao, J. Chen, W. Peng, et al., "Effects of Growth Hormone Over-Expression on Reproduction in the Common Carp *Cyprinus carpio* L.," *General and Comparative Endocrinology* 195 (2014): 47–57, <https://doi.org/10.1016/j.ygcen.2013.10.011>.
109. J. Dai, X. Cui, Z. Zhu, and W. Hu, "Non-Homologous End Joining Plays a Key Role in Transgene Concatemer Formation in Transgenic Zebrafish Embryos," *International Journal of Biological Sciences* 6, no. 7 (2010): 756–768, <https://doi.org/10.7150/ijbs.6.756>.
110. R. A. Dunham, "Transgenic Fish Resistant to Infectious Diseases, Their Risk and Prevention of Escape Into the Environment and Future Candidate Genes for Disease Transgene Manipulation," *Comparative Immunology, Microbiology and Infectious Diseases* 32, no. 2 (2009): 139–161, <https://doi.org/10.1016/j.cimid.2007.11.006>.
111. G. Wu, Y. Sun, and Z. Zhu, "Growth Hormone Gene Transfer in Common Carp," *Aquatic Living Resources* 16, no. 5 (2003): 416–420, [https://doi.org/10.1016/S0990-7440\(03\)00087-1](https://doi.org/10.1016/S0990-7440(03)00087-1).
112. Y. Hong, S. Chen, J. Gui, and M. Scharl, "Retention of the Developmental Pluripotency in Medaka Embryonic Stem Cells After Gene Transfer and Long-Term Drug Selection for Gene Targeting in Fish," *Transgenic Research* 13, no. 1 (2004): 41–50, <https://doi.org/10.1023/b:trag.0000017172.71391.fa>.
113. W. Hu, S. Li, B. Tang, et al., "Antisense for Gonadotropin-Releasing Hormone Reduces Gonadotropin Synthesis and Gonadal Development in Transgenic Common Carp (*Cyprinus carpio*)," *Aquaculture* 271, no. 1 (2007): 498–506, <https://doi.org/10.1016/j.aquaculture.2007.04.075>.
114. W. Hu and Z. Zhu, "Integration Mechanisms of Transgenes and Population Fitness of GH Transgenic Fish," *Science China. Life Sciences* 53, no. 4 (2010): 401–408, <https://doi.org/10.1007/s11427-010-0088-2>.
115. D. Li, C. Fu, Y. Wang, Z. Zhu, and W. Hu, "The Hematological Response to Exhaustive Exercise in 'All-Fish' Growth Hormone Transgenic Common Carp (*Cyprinus carpio* L.)," *Aquaculture* 311 (2011): 263–268, <https://doi.org/10.1016/j.aquaculture.2010.12.002>.
116. C. Zhong, Y. Song, Y. Wang, et al., "Growth Hormone Transgene Effects on Growth Performance Are Inconsistent Among Offspring Derived From Different Homozygous Transgenic Common Carp (*Cyprinus carpio* L.)," *Aquaculture* 356–357 (2012): 404–411, <https://doi.org/10.1016/j.aquaculture.2012.04.019>.
117. C. Zhong, Y. Song, Y. Wang, et al., "Increased Food Intake in Growth Hormone-Transgenic Common Carp (*Cyprinus carpio* L.) May Be Mediated by Upregulating Agouti-Related Protein (AgRP)," *General and Comparative Endocrinology* 192 (2013): 81–88, <https://doi.org/10.1016/j.ygcen.2013.03.024>.
118. G. Chen, J. Huang, J. Jia, et al., "The Food Safety Assessment of All-Female Common Carp (*Cyprinus carpio*) (cyp17a1+/-;XX Genotype) Generated Using Genome Editing Technology," *Food and Chemical Toxicology* 181 (2023): 114103, <https://doi.org/10.1016/j.fct.2023.114103>.
119. P. P. Chiou, M. J. Chen, C. M. Lin, et al., "Production of Homozygous Transgenic Rainbow Trout With Enhanced Disease Resistance," *Marine Biotechnology (New York, N.Y.)* 16, no. 3 (2014): 299–308, <https://doi.org/10.1007/s10126-013-9550-z>.
120. S. J. Du, Z. Gong, G. L. Fletcher, et al., "Growth Enhancement in Transgenic Atlantic Salmon by the Use of an 'All Fish' Chimeric Growth Hormone Gene Construct," *Bio/Technology* 10, no. 2 (1992): 176–181, <https://doi.org/10.1038/nbt0292-176>.
121. E. S. Yaskowiak, M. A. Shears, A. Agarwal-Mawal, and G. L. Fletcher, "Characterization and Multi-Generational Stability of the Growth Hormone Transgene (EO-1 $\alpha$ ) Responsible for Enhanced Growth Rates in Atlantic Salmon," *Transgenic Research* 15, no. 4 (2006): 465–480, <https://doi.org/10.1007/s11248-006-0020-5>.
122. E. H. Ignatz, T. S. Hori, S. Kumar, et al., "RNA-Seq Analysis of the Growth Hormone Transgenic Female Triploid Atlantic Salmon (*Salmo salar*) Hepatic Transcriptome Reveals Broad Temperature-Mediated Effects on Metabolism and Other Biological Processes," *Frontiers in Genetics* 13 (2022): 852165, <https://doi.org/10.3389/fgene.2022.852165>.
123. N. Chang, C. Sun, L. Gao, et al., "Genome Editing With RNA-Guided Cas9 Nuclease in Zebrafish Embryos," *Cell Research* 23, no. 4 (2013): 465–472, <https://doi.org/10.1038/cr.2013.45>.
124. L. Chu, J. Li, Y. Liu, W. Hu, and C. H. Cheng, "Targeted Gene Disruption in Zebrafish Reveals Noncanonical Functions of LH Signaling in Reproduction," *Molecular Endocrinology* 28, no. 11 (2014): 1785–1795, <https://doi.org/10.1210/me.2014-1061>.
125. Y. Doyon, J. M. McCammon, J. C. Miller, et al., "Heritable Targeted Gene Disruption in Zebrafish Using Designed Zinc-Finger Nucleases," *Nature Biotechnology* 26, no. 6 (2008): 702–708, <https://doi.org/10.1038/nbt1409>.
126. A. Hruscha, P. Krawitz, A. Rechenberg, et al., "Efficient CRISPR/Cas9 Genome Editing With Low Off-Target Effects in Zebrafish," *Development* 140, no. 24 (2013): 4982–4987, <https://doi.org/10.1242/dev.099085>.



127. W. Y. Hwang, Y. Fu, D. Reyon, et al., "Efficient Genome Editing in Zebrafish Using a CRISPR-Cas System," *Nature Biotechnology* 31, no. 3 (2013): 227–229, <https://doi.org/10.1038/nbt.2501>.
128. Y. Shu, Q. Lou, Z. Dai, et al., "The Basal Function of Teleost Prolactin as a Key Regulator on Ion Uptake Identified With Zebrafish Knockout Models," *Scientific Reports* 6, no. 1 (2016): 18597, <https://doi.org/10.1038/srep18597>.
129. H. Tang, Y. Liu, D. Luo, et al., "The Kiss/Kissr Systems Are Dispensable for Zebrafish Reproduction: Evidence From Gene Knockout Studies," *Endocrinology* 156, no. 2 (2015): 589–599, <https://doi.org/10.1210/en.2014-1204>.
130. S. Ansai, K. Inohaya, Y. Yoshiura, et al., "Design, Evaluation, and Screening Methods for Efficient Targeted Mutagenesis With Transcription Activator-Like Effector Nucleases in Medaka," *Development, Growth and Differentiation* 56, no. 1 (2014): 98–107, <https://doi.org/10.1111/dgd.12104>.
131. S. Ansai, T. Sakuma, T. Yamamoto, et al., "Efficient Targeted Mutagenesis in Medaka Using Custom-Designed Transcription Activator-Like Effector Nucleases," *Genetics* 193, no. 3 (2013): 739–749, <https://doi.org/10.1534/genetics.112.147645>.
132. Y. A. Chiang, M. Kinoshita, S. Maekawa, et al., "TALENs-Mediated Gene Disruption of Myostatin Produces a Larger Phenotype of Medaka With an Apparently Compromised Immune System," *Fish and Shellfish Immunology* 48 (2016): 212–220, <https://doi.org/10.1016/j.fsi.2015.11.016>.
133. K. Feng, H. Luo, Y. Li, et al., "High Efficient Gene Targeting in Rice Field Eel *Monopterus albus* by Transcription Activator-Like Effector Nucleases," *Science Bulletin* 62, no. 3 (2017): 162–164, <https://doi.org/10.1016/j.scib.2017.01.018>.
134. J. Chen, W. Wang, Z. Tian, et al., "Efficient Gene Transfer and Gene Editing in Sterlet (*Acipenser ruthenus*)," *Frontiers in Genetics* 9 (2018): 117, <https://doi.org/10.3389/fgene.2018.00117>.
135. Z. Qin, Y. Li, B. Su, et al., "Editing of the Luteinizing Hormone Gene to Sterilize Channel Catfish, *Ictalurus punctatus*, Using a Modified Zinc Finger Nuclease Technology With Electroporation," *Marine Biotechnology (New York, N.Y.)* 18, no. 2 (2016): 255–263, <https://doi.org/10.1007/s10126-016-9687-7>.
136. V. Chakrapani, S. K. Patra, R. P. Panda, K. D. Rasal, P. Jayasankar, and H. K. Barman, "Establishing Targeted Carp TLR22 Gene Disruption via Homologous Recombination Using CRISPR/Cas9," *Developmental and Comparative Immunology* 61 (2016): 242–247, <https://doi.org/10.1016/j.dci.2016.04.009>.
137. Z. Zhong, P. Niu, M. Wang, et al., "Targeted Disruption of sp7 and Myostatin With CRISPR-Cas9 Results in Severe Bone Defects and More Muscular Cells in Common Carp," *Scientific Reports* 6, no. 1 (2016): 22953, <https://doi.org/10.1038/srep22953>.
138. Y. Ou, H. Li, J. Li, et al., "Formation of Different Polyploids Through Disrupting Meiotic Crossover Frequencies Based on cntd1 Knockout in Zebrafish," *Molecular Biology and Evolution* 41, no. 3 (2024): msae047, <https://doi.org/10.1093/molbev/msae047>.
139. H. Xu, G. Tong, T. Yan, et al., "Transcriptomic Analysis Provides Insights to Reveal the bmp6 Function Related to the Development of Intermuscular Bones in Zebrafish," *Frontiers in Cell and Development Biology* 10 (2022): 821471, <https://doi.org/10.3389/fcell.2022.821471>.
140. G. Tang, W. Lv, Z. Sun, et al., "Heritability and Quantitative Trait Locus Analyses of Intermuscular Bones in Mirror Carp (*Cyprinus carpio*)," *Aquaculture* 515 (2020): 734601, <https://doi.org/10.1016/j.aquaculture.2019.734601>.
141. X.-D. Wang, C.-H. Nie, and Z.-X. Gao, "Research Progress on Molecular Regulation Mechanism and Genetic Selection of Intermuscular Bones in Teleosts," *Acta Hydrobiologica Sinica* 45, no. 3 (2021): 680–691, <https://doi.org/10.7541/2021.2020.092>.
142. C.-H. Nie, A. W. S. Hilsdorf, S.-M. Wan, and Z.-X. Gao, "Understanding the Development of Intermuscular Bones in Teleost: Status and Future Directions for Aquaculture," *Reviews in Aquaculture* 12, no. 2 (2020): 759–772, <https://doi.org/10.1111/raq.12348>.
143. X.-M. Xiong, W.-J. Huang, Q. Dong, D.-Y. Zhang, S.-M. Wan, and Z.-X. Gao, "Genetic Parameter Estimates for Intermuscular Bone Trait in Grass Carp (*Ctenopharyngodon idella*)," *Aquaculture* 563 (2023): 739011, <https://doi.org/10.1016/j.aquaculture.2022.739011>.
144. Y. Kuang, X. Zheng, D. Cao, et al., "Generate a New Crucian Carp (*Carassius auratus*) Strain Without Intermuscular Bones by Knocking out bmp6," *Aquaculture* 569 (2023): 739407, <https://doi.org/10.1016/j.aquaculture.2023.739407>.
145. Q. Dong, C.-H. Nie, Y.-M. Wu, et al., "Generation of Blunt Snout Bream Without Intermuscular Bones by Runx2b Gene Mutation," *Aquaculture* 567 (2023): 739263, <https://doi.org/10.1016/j.aquaculture.2023.739263>.
146. C. H. Nie, S. M. Wan, Y. L. Chen, et al., "Single-Cell Transcriptomes and Runx2b(–/–) Mutants Reveal the Genetic Signatures of Intermuscular Bone Formation in Zebrafish," *National Science Review* 9, no. 11 (2022): nwac152, <https://doi.org/10.1093/nsr/nwac152>.
147. W. He, Q. Qin, S. Liu, et al., "Organization and Variation Analysis of 5S rDNA in Different Ploidy-Level Hybrids of Red Crucian Carp × Topmouth Culter," *PLoS One* 7, no. 6 (2012): e38976, <https://doi.org/10.1371/journal.pone.0038976>.
148. W. Huang, Q. Qin, H. Yang, et al., "Formation of Diploid and Triploid Hybrid Groupers (Hybridization of *Epinephelus coioides* ♀ × *Epinephelus lanceolatus* ♂) and Their 5S Gene Analysis," *BMC Genetics* 17, no. 1 (2016): 136, <https://doi.org/10.1186/s12863-016-0443-9>.
149. J. Hu, S. Liu, J. Xiao, et al., "Characteristics of Diploid and Triploid Hybrids Derived From Female *Megalobrama amblycephala* Yih × Male *Xenocypris davidi* Bleeker," *Aquaculture* 364–365 (2012): 157–164, <https://doi.org/10.1016/j.aquaculture.2012.08.025>.
150. S. Wang, X. Ye, Y. Wang, et al., "A New Type of Homodiploid Fish Derived From the Interspecific Hybridization of Female Common Carp × Male Blunt Snout Bream," *Scientific Reports* 7, no. 1 (2017): 4189, <https://doi.org/10.1038/s41598-017-04582-z>.
151. S. Liu, Y. Liu, G. Zhou, et al., "The Formation of Tetraploid Stocks of Red Crucian Carp × Common Carp Hybrids as an Effect of Interspecific Hybridization," *Aquaculture* 192, no. 2 (2001): 171–186, [https://doi.org/10.1016/S0044-8486\(00\)00451-8](https://doi.org/10.1016/S0044-8486(00)00451-8).
152. J. Xiao, F. Hu, K. Luo, W. Li, and S. Liu, "Unique Nucleolar Dominance Patterns in Distant Hybrid Lineage Derived From *Megalobrama amblycephala* × *Culter alburnus*," *BMC Genetics* 17, no. 1 (2016): 150, <https://doi.org/10.1186/s12863-016-0457-3>.
153. Y. Wang, C. Yang, K. Luo, et al., "The Formation of the Goldfish-Like Fish Derived From Hybridization of Female Koi Carp × Male Blunt Snout Bream," *Frontiers in Genetics* 9 (2018): 437, <https://doi.org/10.3389/fgene.2018.00437>.
154. Z. Zhang, J. Chen, L. Li, et al., "Research Advances in Animal Distant Hybridization," *Science China. Life Sciences* 57, no. 9 (2014): 889–902, <https://doi.org/10.1007/s11427-014-4707-1>.
155. Q. Liu, S. Wang, C. Tang, et al., "The Research Advances in Distant Hybridization and Gynogenesis in Fish," *Reviews in Aquaculture* 17, no. 1 (2025): e12972, <https://doi.org/10.1111/raq.12972>.
156. H. Zhong, H. Chen, M. Liu, et al., "Improved Traits of Proximate Composition, Liver Antioxidant Capacity and Feeding Habits in Diploid Hybrids From Female *Micropterus salmoides* × Male *Lepomis cyanellus*," *Aquaculture* 587 (2024): 740853, <https://doi.org/10.1016/j.aquaculture.2024.740853>.
157. C. Jie, L. Mi, L. Shengnan, et al., "A Comparative Study of Distant Hybridization in Plants and Animals," *Science China Life Sciences* 61, no. 3 (2018): 285–309, <https://doi.org/10.1007/s11427-017-9094-2>.



158. B. M. McCluskey, P. Batzel, and J. H. Postlethwait, "The Hybrid History of Zebrafish," *G3 (Bethesda)* 15, no. 2 (2025): jkae299, <https://doi.org/10.1093/g3journal/jkae299>.
159. S. Liu, "Distant Hybridization Leads to Different Ploidy Fishes," *Science China Life Sciences* 53, no. 4 (2010): 416–425, <https://doi.org/10.1007/s11427-010-0057-9>.
160. Y. Wang, X.-Y. Li, W.-J. Xu, et al., "Comparative Genome Anatomy Reveals Evolutionary Insights Into a Unique Amphitriploid Fish," *Nature Ecology and Evolution* 6, no. 9 (2022): 1354–1366, <https://doi.org/10.1038/s41559-022-01813-z>.
161. S. Wang, N. Jiao, L. Zhao, et al., "Evidence for the Paternal Mitochondrial DNA in the Crucian Carp-Like Fish Lineage With Hybrid Origin," *Science China Life Sciences* 63, no. 1 (2020): 102–115, <https://doi.org/10.1007/s11427-019-9528-1>.
162. M. Ou, J. Zhao, Q. Luo, et al., "Characteristics of Hybrids Derived From *Channa argus* ♀ × *Channa maculata* ♂," *Aquaculture* 492 (2018): 349–356, <https://doi.org/10.1016/j.aquaculture.2018.04.038>.
163. Q. Liu, Y. Qi, Q. Liang, et al., "The Chimeric Genes in the Hybrid Lineage of *Carassius auratus* Cuvieri (♀) × *Carassius auratus* Red Var. (♂)," *Science China Life Sciences* 61, no. 9 (2018): 1079–1089, <https://doi.org/10.1007/s11427-017-9306-7>.
164. D. Gong, L. Xu, Q. Liu, et al., "A New Type of Hybrid Bream Derived From a Hybrid Lineage of *Megalobrama amblycephala* (♀) × *Culter alburnus* (♂)," *Aquaculture* 534 (2021): 736194, <https://doi.org/10.1016/j.aquaculture.2020.736194>.
165. D. I. Bolnick, "Hybridization and Speciation in Centrarchids," in *Centrarchid Fishes* (Wiley-Blackwell, 2009), 39–69.
166. W. Childers, "Hybridization of Four Species of Sunfishes (Centrarchidae)," *Illinois Natural History Survey Bulletin* 29 (1967): 159–214, <https://doi.org/10.21900/j.inhs.v29.165>.
167. P. Berrebi, G. Cattaneo-Berrebi, and N. Le Brun, "Natural Hybridization of Two Species of Tetraploid Barbels: *Barbus meridionalis* and *Barbus barbus* (Osteichthyes, Cyprinidae) in Southern France," *Biological Journal of the Linnean Society* 48, no. 4 (1993): 319–333, [https://doi.org/10.1016/0024-4066\(93\)90003-7](https://doi.org/10.1016/0024-4066(93)90003-7).
168. T. E. Dowling and B. D. DeMarais, "Evolutionary Significance of Introgressive Hybridization in Cyprinid Fishes," *Nature* 362, no. 6419 (1993): 444–446, <https://doi.org/10.1038/362444a0>.
169. G. Hulata, "A Review of Genetic Improvement of the Common Carp (*Cyprinus carpio* L.) and Other Cyprinids by Crossbreeding, Hybridization and Selection," *Aquaculture* 129, no. 1 (1995): 143–155, [https://doi.org/10.1016/0044-8486\(94\)00244-I](https://doi.org/10.1016/0044-8486(94)00244-I).
170. A. Simková, M. Dávidová, I. Papoušek, and L. Vetešník, "Does Interspecies Hybridization Affect the Host Specificity of Parasites in Cyprinid Fish?," *Parasites and Vectors* 6 (2013): 95, <https://doi.org/10.1186/1756-3305-6-95>.
171. A. Šimková, L. Vojtek, K. Halačka, P. Hyršl, and L. Vetešník, "The Effect of Hybridization on Fish Physiology, Immunity and Blood Biochemistry: A Case Study in Hybridizing *Cyprinus carpio* and *Carassius gibelio* (Cyprinidae)," *Aquaculture* 435 (2015): 381–389, <https://doi.org/10.1016/j.aquaculture.2014.10.021>.
172. Y. Zhou, L. Ren, J. Xiao, et al., "Global Transcriptional and miRNA Insights Into Bases of Heterosis in Hybridization of Cyprinidae," *Scientific Reports* 5, no. 1 (2015): 13847, <https://doi.org/10.1038/srep13847>.
173. R. O. Lowe-McConnell, "Ecology and Evolution of Poeciliid Fishes," *Zoological Journal of the Linnean Society* 166, no. 3 (2012): 688, <https://doi.org/10.1111/j.1096-3642.2012.00843.x>.
174. J. M. Blanc and B. Chevassus, "Interspecific Hybridization of Salmonid Fish," *Aquaculture* 18, no. 1 (1979): 21–34, [https://doi.org/10.1016/0044-8486\(79\)90097-8](https://doi.org/10.1016/0044-8486(79)90097-8).
175. B. Chevassus, "Hybridization in Salmonids: Results and Perspectives," *Aquaculture* 17, no. 2 (1979): 113–128, [https://doi.org/10.1016/0044-8486\(79\)90047-4](https://doi.org/10.1016/0044-8486(79)90047-4).
176. D.-D. Guo, F. Liu, B.-L. Niu, et al., "Establishment of Diploid Hybrid Strains Derived From Female *Larimichthys crocea* × Male *Larimichthys polyactis* and Transmission of Parental mtDNA in Hybrid Progenies," *Aquaculture* 561 (2022): 738693, <https://doi.org/10.1016/j.aquaculture.2022.738693>.
177. W. Huang, Q. Liu, J. Xie, et al., "Characterization of Triploid Hybrid Groupers From Interspecies Hybridization (*Epinephelus coioides* ♀ × *Epinephelus lanceolatus* ♂)," *Aquaculture Research* 47, no. 7 (2016): 2195–2204, <https://doi.org/10.1111/are.12672>.
178. L. Xiao, D. Wang, Y. Guo, et al., "Comparative Transcriptome Analysis of Diploid and Triploid Hybrid Groupers (*Epinephelus coioides* ♀ × *E. lanceolatus* ♂) Reveals the Mechanism of Abnormal Gonadal Development in Triploid Hybrids," *Genomics* 111, no. 3 (2019): 251–259, <https://doi.org/10.1016/j.ygeno.2018.11.010>.
179. L. Fantini-Hoag, T. Hanson, F. Kubitzka, J. A. Povh, R. A. C. Corrêa Filho, and J. Chappell, "Growth Performance and Economic Analysis of Hybrid Catfish (Channel Catfish *Ictalurus punctatus* ♀ × Blue Catfish, *I. furcatus* ♂) and Channel Catfish (*I. punctatus*) Produced in Floating in-Pond Raceway System," *Aquaculture Reports* 23 (2022): 101065, <https://doi.org/10.1016/j.aqrep.2022.101065>.
180. N. G. Chatakondi, R. D. Yant, and R. A. Dunham, "Effect of Paternal Blue Catfish Strain Effects on Hatchery Fry Production and Performance of Channel Catfish × Blue Catfish F1 Hybrid Fry Production and Fingerling Performance Under Commercial Conditions," *North American Journal of Aquaculture* 78, no. 4 (2016): 301–306, <https://doi.org/10.1080/15222055.2016.1185065>.
181. L. Zhou, Y. Wang, and J.-F. Gui, "Genetic Evidence for Gonochoristic Reproduction in Gynogenetic Silver Crucian Carp (*Carassius auratus* Gibelio Bloch) as Revealed by RAPD Assays," *Journal of Molecular Evolution* 51, no. 5 (2000): 498–506, <https://doi.org/10.1007/s002390010113>.
182. J. Yan, S. Liu, Y. Sun, C. Zhang, K. Luo, and Y. Liu, "RAPD and Microsatellite Analysis of Diploid Gynogens From Allotetraploid Hybrids of Red Crucian Carp (*Carassius auratus*) × Common Carp (*Cyprinus Carpio*)," *Aquaculture* 243, no. 1 (2005): 49–60, <https://doi.org/10.1016/j.aquaculture.2004.09.025>.
183. Q. Liu, X. Zhang, J. Liu, et al., "A New Type of Allodiploid Hybrids Derived From Female *Megalobrama amblycephala* × Male *Gobiocypris rarus*," *Frontiers in Genetics* 12 (2021): 685914, <https://doi.org/10.3389/fgene.2021.685914>.
184. S. Liu, W. Duan, M. Tao, et al., "Establishment of the Diploid Gynogenetic Hybrid Clonal Line of Red Crucian Carp × Common Carp," *Science in China Series C: Life Sciences* 50, no. 2 (2007): 186–193, <https://doi.org/10.1007/s11427-007-0032-2>.
185. J. Gui and L. Zhou, "Genetic Basis and Breeding Application of Clonal Diversity and Dual Reproduction Modes in Polyploid *Carassius auratus* Gibelio," *Science China. Life Sciences* 53, no. 4 (2010): 409–415, <https://doi.org/10.1007/s11427-010-0092-6>.
186. J. Liu and G. Yang, "Changes in Copper Content of Allogynogenetic Silver Crucian Carp After Application of Copper Sulfate to Fishponds," *Israeli Journal of Aquaculture – Bamideg* 61 (2009): 351–355, <https://doi.org/10.46989/001c.20563>.
187. D. Wang, H. Mao, J. Peng, X. Li, L. Zhou, and J. Gui, "Discovery of a Male-Biased Mutant Family and Identification of a Male-Specific SCAR Marker in Gynogenetic Gibel Carp *Carassius auratus* Gibelio," *Progress in Natural Science* 19, no. 11 (2009): 1537–1544, <https://doi.org/10.1016/j.pnsc.2009.04.008>.
188. S. Wang, C. Tang, M. Tao, et al., "Establishment and Application of Distant Hybridization Technology in Fish," *Science China Life Sciences* 62, no. 1 (2019): 22–45, <https://doi.org/10.1007/s11427-018-9408-x>.

189. S. Liu, Q. Qin, Y. Wang, et al., "Evidence for the Formation of the Male Gynogenetic Fish," *Marine Biotechnology (New York, N.Y.)* 12, no. 2 (2010): 160–172, <https://doi.org/10.1007/s10126-009-9219-9>.
190. Q. Qin, C. Wang, Y. Zhou, et al., "Rapid Genomic and Epigenetic Alterations in Gynogenetic *Carassius auratus* Red Var. Derived From Distant Hybridization," *Marine Biotechnology (New York, N.Y.)* 22, no. 3 (2020): 433–442, <https://doi.org/10.1007/s10126-020-09963-6>.
191. C. Wu, X. Huang, F. Hu, et al., "Production of Diploid Gynogenetic Grass Carp and Triploid Hybrids Derived From the Distant Hybridization of Female Grass Carp and Male Topmouth Culter," *Aquaculture* 504 (2019): 462–470, <https://doi.org/10.1016/j.aquaculture.2018.12.056>.
192. Y. Wang, J. Yao, A. Liao, et al., "The Formation of Hybrid Fish Derived From Hybridization of *Megalobrama amblycephala* (♀) × *Siniperca chuatsi* (♂)," *Aquaculture* 548 (2022): 737547, <https://doi.org/10.1016/j.aquaculture.2021.737547>.
193. S. Liu, Y. Wang, S. Wang, Y. Zhou, C. Zhang, and M. Tao, "The Formation and Biological Characteristics of Different Ploidy Fishes Derived From Common Carp × Blunt Snout Bream," in *Fish Distant Hybridization* (Springer Nature Singapore, 2022), 233–270.
194. J. Li, Y. Pan, Q. Xing, et al., "Trait Segregation and Growth Characteristics of the F2 Generation Hybrids Between *Crassostrea* *Dianbaiensis* and *C. iredalei*," *Aquaculture International* 33, no. 4 (2025): 274, <https://doi.org/10.1007/s10499-025-01951-8>.
195. C. Krueger and B. May, "Ecological and Genetic Effects of Salmonid Introductions in North America," *Canadian Journal of Fisheries and Aquatic Sciences* 48 (1991): 66–77, <https://doi.org/10.1139/f91-305>.
196. J. M. Yáñez, J. M. Yáñez, S. eNewman, and R. D. Houston, "Genomics in Aquaculture to Better Understand Species Biology and Accelerate Genetic Progress," *Frontiers in Genetics* 6 (2015): 128, <https://doi.org/10.3389/fgene.2015.00128>.
197. T. A. Delomas and K. Dabrowski, "Zebrafish Embryonic Development Is Induced by Carp Sperm," *Biology Letters* 12, no. 11 (2016): 20160628, <https://doi.org/10.1098/rsbl.2016.0628>.
198. R. Rożyński, M. Kuciński, S. Dobosz, and K. Ocalewicz, "Successful Application of UV-Irradiated Rainbow Trout (*Oncorhynchus mykiss*) Spermatozoa to Induce Gynogenetic Development of the European Grayling (*Thymallus thymallus*)," *Aquaculture* 574 (2023): 739720, <https://doi.org/10.1016/j.aquaculture.2023.739720>.
199. R. M. Bertolini, L. S. López, B. dos Santos Machado, et al., "Gynogenesis in the Spotted Catfish *Pimelodus maculatus* Using Heterologous Gametes From the Yellowtail-Tetra (*Astyanax altiparanae*): A Key Point to Achieve Monosex Female Populations in Neotropical Catfishes," *Aquaculture International* 33, no. 2 (2025): 109, <https://doi.org/10.1007/s10499-024-01785-w>.
200. J. Komen, G. F. Wiegertjes, V. J. T. van Ginneken, E. H. Eding, and C. J. J. Richter, "Gynogenesis in Common Carp (*Cyprinus carpio* L.). III. The Effects of Inbreeding on Gonadal Development of Heterozygous and Homozygous Gynogenetic Offspring," *Aquaculture* 104, no. 1 (1992): 51–66, [https://doi.org/10.1016/0044-8486\(92\)90137-A](https://doi.org/10.1016/0044-8486(92)90137-A).
201. D. Fopp-Bayat, R. Kolman, and P. Woznicki, "Induction of Meiotic Gynogenesis in Sterlet (*Acipenser ruthenus*) Using UV-Irradiated Bester Sperm," *Aquaculture* 264, no. 1 (2007): 54–58, <https://doi.org/10.1016/j.aquaculture.2006.12.006>.
202. A. J. Morgan, R. Murashige, C. A. Woolridge, et al., "Effective UV Dose and Pressure Shock for Induction of Meiotic Gynogenesis in Southern Flounder (*Paralichthys lethostigma*) Using Black Sea Bass (*Centropomus striata*) Sperm," *Aquaculture* 259, no. 1 (2006): 290–299, <https://doi.org/10.1016/j.aquaculture.2006.05.045>.
203. F. Piferrer, R. M. Cal, C. Gómez, B. Álvarez-Blázquez, J. Castro, and P. Martínez, "Induction of Gynogenesis in the Turbot (*Scophthalmus maximus*): Effects of UV Irradiation on Sperm Motility, the Hertwig Effect and Viability During the First 6 Months of Age," *Aquaculture* 238, no. 1 (2004): 403–419, <https://doi.org/10.1016/j.aquaculture.2004.05.009>.
204. G. H. Thorgaard, P. D. Scheerer, W. K. Hershberger, and J. M. Myers, "Androgenetic Rainbow Trout Produced Using Sperm From Tetraploid Males Show Improved Survival," *Aquaculture* 85, no. 1 (1990): 215–221, [https://doi.org/10.1016/0044-8486\(90\)90021-E](https://doi.org/10.1016/0044-8486(90)90021-E).
205. T. Fujimoto, G. S. Yasui, M. Hayakawa, S. Sakao, E. Yamaha, and K. Arai, "Reproductive Capacity of Neo-Tetraploid Loaches Produced Using Diploid Spermatozoa From a Natural Tetraploid Male," *Aquaculture* 308 (2010): S133–S139, <https://doi.org/10.1016/j.aquaculture.2010.04.029>.
206. F. Lin and K. Dabrowski, "Androgenesis and Homozygous Gynogenesis in Muskellunge (*Esox masquinongy*): Evaluation Using Flow Cytometry," *Molecular Reproduction and Development* 49, no. 1 (1998): 10–18, [https://doi.org/10.1002/\(sici\)1098-2795\(199801\)49:1<10::Aid-mrd2>3.0.Co;2-r](https://doi.org/10.1002/(sici)1098-2795(199801)49:1<10::Aid-mrd2>3.0.Co;2-r).
207. Q. Li and A. Kijima, "Microsatellite Analysis of Gynogenetic Families in the Pacific Oyster, *Crassostrea gigas*," *Journal of Experimental Marine Biology and Ecology* 331, no. 1 (2005): 1–8, <https://doi.org/10.1016/j.jembe.2005.09.009>.
208. P. Wu, Y. Zeng, Q. Qin, et al., "Formation and Identification of Artificial Gynogenetic Mandarin Fish (*Siniperca chuatsi*) Induced by Inactivated Sperm of Largemouth Bass (*Micropterus salmoides*)," *Aquaculture* 577 (2023): 739969, <https://doi.org/10.1016/j.aquaculture.2023.739969>.
209. Y. Wang, A. m. Liao, C. Geng, et al., "The Formation and Study of Allogynogenesis *Hemibarbus maculatus* Bleeker," *Reproduction and Breeding* 3, no. 1 (2023): 1–7, <https://doi.org/10.1016/j.repbre.2022.12.002>.
210. W. Li, Z. Zhou, X. Tian, et al., "Gynogenetic *Cirrhinus mrigala* Produced Using Irradiated Sperm of *Cyprinus carpio* Exhibit Better Cold Tolerance," *Reproduction and Breeding* 3, no. 1 (2023): 8–16, <https://doi.org/10.1016/j.repbre.2023.01.001>.
211. H. Zhong, Y. Sun, M. Liu, et al., "Induction of Diploid Gynogenesis in *Micropterus salmoides* Using Irradiated Heterogeneous Sperm From *Siniperca chuatsi*," *Aquaculture* 590 (2024): 741021, <https://doi.org/10.1016/j.aquaculture.2024.741021>.
212. Y. D. Sun, C. Zhang, S. J. Liu, M. Tao, C. Zeng, and Y. Liu, "Induction of Gynogenesis in Japanese Crucian Carp (*Carassius cuvieri*)," *Yi Chuan Xue Bao* 33, no. 5 (2006): 405–412, [https://doi.org/10.1016/s0379-4172\(06\)60067-x](https://doi.org/10.1016/s0379-4172(06)60067-x).
213. K. Luo, J. Xiao, S. Liu, et al., "Massive Production of All-Female Diploids and Triploids in the Crucian Carp," *International Journal of Biological Sciences* 7, no. 4 (2011): 487–495, <https://doi.org/10.7150/ijbs.7.487>.
214. D. Gong, L. Xu, C. Wu, et al., "Two Types of Gynogenetic Blunt Snout Bream Derived From Different Sperm," *Aquaculture* 511 (2019): 734250, <https://doi.org/10.1016/j.aquaculture.2019.734250>.
215. J. Xiao, T. M. Zou, L. Chen, et al., "Microsatellite Analysis of Different Ploidy Offspring of Artificial Gynogenesis in *Cyprinus carpio*," *Journal of Fish Biology* 78, no. 1 (2011): 150–165, <https://doi.org/10.1111/j.1095-8649.2010.02848.x>.
216. Z. Mao, Y. Fu, S. Wang, et al., "Further Evidence for Paternal DNA Transmission in Gynogenetic Grass Carp," *Science China. Life Sciences* 63, no. 9 (2020): 1287–1296, <https://doi.org/10.1007/s11427-020-1698-x>.
217. A. Liao, S. Zhang, Q. Yu, et al., "Formation and Characterization of Artificial Gynogenetic Northern Snakehead (*Channa argus*) Induced by Inactivated Sperm of Mandarin Fish (*Siniperca chuatsi*)," *Aquaculture* 595 (2025): 741488, <https://doi.org/10.1016/j.aquaculture.2024.741488>.
218. S. Wang, P. Zhou, X. Huang, et al., "The Establishment of an Autotetraploid Fish Lineage Produced by Female Allotetraploid Hybrids × Male Homodiploid Hybrids Derived From *Cyprinus carpio*

- (♀) × *Megalobrama amblycephala* (♂),” *Aquaculture* 515 (2020): 734583, <https://doi.org/10.1016/j.aquaculture.2019.734583>.
219. Q. Qin, Y. Wang, J. Wang, et al., “The Autotetraploid Fish Derived From Hybridization of *Carassius auratus* Red Var. (Female) × *Megalobrama amblycephala* (Male),” *Biology of Reproduction* 91, no. 4 (2014): 93, <https://doi.org/10.1095/biolreprod.114.122283>.
220. S. Liu, Q. Qin, J. Xiao, et al., “The Formation of the Polyploid Hybrids From Different Subfamily Fish Crossings and Its Evolutionary Significance,” *Genetics* 176, no. 2 (2007): 1023–1034, <https://doi.org/10.1534/genetics.107.071373>.
221. Q. Liu, Y. Fan, Z. Xiong, et al., “Unique Nucleolar Dominance Patterns in Different Ploidy Hybrid Lineages Derived From *Cyprinus carpio* (♀) × *Megalobrama amblycephala* (♂),” *Aquaculture* 576 (2023): 739753, <https://doi.org/10.1016/j.aquaculture.2023.739753>.
222. S. Yuandong, T. Min, L. Shaojun, et al., “Induction of Gynogenesis in Red Crucian Carp Using Spermatozoa of Blunt Snout Bream,” *Progress in Natural Science* 17, no. 2 (2007): 163–167, <https://doi.org/10.1080/10020070612331343241>.
223. Q. Liu, J. Xiao, K. Luo, Y. Zhang, and Y. Wang, “Genetic, Gonadal Development and Shape Characteristics Researches of Gynogenetic Orange Ornamental Carp (*Cyprinus carpio* L.),” *Journal of Fisheries of China* 37 (2013): 390.
224. P. Wu, Y. Zeng, Q. Qin, et al., “Comparative Analysis of the Texture, Composition, Antioxidant Capacity and Nutrients of Natural Gynogenesis Blunt Snout Bream and Its Parent Muscle,” *Reproduction and Breeding* 2, no. 4 (2022): 149–155, <https://doi.org/10.1016/j.repbre.2022.12.001>.
225. J. G. Stanley, “Production of Hybrid, Androgenetic, and Gynogenetic Grass Carp and Carp,” *Transactions of the American Fisheries Society* 105 (1976): 10–16.
226. S. Liu, F. Hu, Y. Zhou, C. Wu, H. Zhong, and K. Luo, “The Formation and Biological Characteristics of the Different Ploidy Fish Derived From the Hybridization of Japanese White Crucian Carp × Blunt Snout Bream,” in *Fish Distant Hybridization* (Springer Nature Singapore, 2022), 189–207.
227. F. Hu, C. Wu, Y. Zhou, et al., “Production of Androgenetic, Triploid and Tetraploid Hybrids From the Interspecific Hybridization of Female Japanese Crucian Carp and Male Blunt Snout Bream,” *Aquaculture* 491 (2018): 50–58, <https://doi.org/10.1016/j.aquaculture.2018.03.014>.
228. K. Arai, H. Onozato, and F. Yamazaki, “Artificial Androgenesis Induced With Gamma Irradiation in Masu Salmon, *Oncorhynchus masou*,” *Bulletin of Fisheries Sciences, Hokkaido University* 30 (1979): 181–186.
229. K. Araki, H. Shinma, H. Nagoya, I. Nakayama, and H. Onozato, “Androgenetic Diploids of Rainbow Trout (*Oncorhynchus mykiss*) Produced by Fused Sperm,” *Canadian Journal of Fisheries and Aquatic Sciences* 52 (2011): 892–896, <https://doi.org/10.1139/f95-089>.
230. I. Babiak, S. Dobosz, K. Goryczko, H. Kuzminski, P. Brzuzan, and S. Ciesielski, “Androgenesis in Rainbow Trout Using Cryopreserved Spermatozoa: The Effect of Processing and Biological Factors,” *Theriogenology* 57, no. 4 (2002): 1229–1249, [https://doi.org/10.1016/S0093-691X\(02\)00631-3](https://doi.org/10.1016/S0093-691X(02)00631-3).
231. Y. K. Nam, Y. S. Cho, H. J. Cho, and D. S. Kim, “Accelerated Growth Performance and Stable Germ-Line Transmission in Androgenetically Derived Homozygous Transgenic Mud Loach, *Misgurnus mizolepis*,” *Aquaculture* 209, no. 1 (2002): 257–270, [https://doi.org/10.1016/S0044-8486\(01\)00730-X](https://doi.org/10.1016/S0044-8486(01)00730-X).
232. J. E. Parsons and G. H. Thorgaard, “Production of Androgenetic Diploid Rainbow Trout,” *Journal of Heredity* 76, no. 3 (1985): 177–181, <https://doi.org/10.1093/oxfordjournals.jhered.a110060>.
233. K. Arai, M. Ikeno, and R. Suzuki, “Production of Androgenetic Diploid Loach *Misgurnus anguillicaudatus* Using Spermatozoa of Natural Tetraploids,” *Aquaculture* 137, no. 1 (1995): 131–138, [https://doi.org/10.1016/0044-8486\(95\)01106-4](https://doi.org/10.1016/0044-8486(95)01106-4).
234. W. Duan, Q. Qin, S. Chen, et al., “The Formation of Improved Tetraploid Population of Red Crucian Carp × Common Carp Hybrids by Androgenesis,” *Science in China Series C: Life Sciences* 50, no. 6 (2007): 753–761, <https://doi.org/10.1007/s11427-007-0090-5>.
235. S. Liu, C. Zhang, Y. Zhou, et al., “The Gynogenesis and Androgenesis of the Diploid Gametes Derived From the Allotetraploid Fish,” in *Fish Distant Hybridization* (Springer Nature Singapore, 2022), 103–143.
236. H. Zhang, S. Liu, C. Zhang, et al., “Induced Gynogenesis in Grass Carp (*Ctenopharyngodon idellus*) Using Irradiated Sperm of Allotetraploid Hybrids,” *Marine Biotechnology* 13, no. 5 (2011): 1017–1026, <https://doi.org/10.1007/s10126-011-9365-8>.
237. Z. Liu, C. Wang, and B. Guo, “Biodiversity—The Cornerstone of Sustainable Aquaculture Development: Insights From the Breeding of Approved Fish Varieties for Aquaculture From 1996 to 2024 in China,” *Reviews in Aquaculture* 17, no. 2 (2025): e70003, <https://doi.org/10.1111/raq.70003>.
238. H. Komen and G. H. Thorgaard, “Androgenesis, Gynogenesis and the Production of Clones in Fishes: A Review,” *Aquaculture* 269, no. 1 (2007): 150–173, <https://doi.org/10.1016/j.aquaculture.2007.05.009>.
239. H. L. Kincaid, “Inbreeding in Fish Populations Used for Aquaculture,” *Aquaculture* 33, no. 1 (1983): 215–227, [https://doi.org/10.1016/0044-8486\(83\)90402-7](https://doi.org/10.1016/0044-8486(83)90402-7).
240. A. Skaarud, J. Woolliams, and H. Gjøen, “Strategies for Controlling Inbreeding in Fish Breeding Programs; an Applied Approach Using Optimum Contribution (OC) Procedures,” *Aquaculture* 311 (2011): 110–114, <https://doi.org/10.1016/j.aquaculture.2010.11.023>.
241. H. Manan, A. B. Noor Hidayati, N. A. Lyana, et al., “A Review of Gynogenesis Manipulation in Aquatic Animals,” *Aquaculture and Fisheries* 7, no. 1 (2022): 1–6, <https://doi.org/10.1016/j.aaf.2020.11.006>.
242. L. Ren, X. Gao, J. Cui, et al., “Symmetric Subgenomes and Balanced Homoeolog Expression Stabilize the Establishment of Allopolyploidy in Cyprinid Fish,” *BMC Biology* 20, no. 1 (2022): 200, <https://doi.org/10.1186/s12915-022-01401-4>.
243. L. Ren, W. Li, Q. Qin, et al., “The Subgenomes Show Asymmetric Expression of Alleles in Hybrid Lineages of *Megalobrama amblycephala* × *Culter alburnus*,” *Genome Research* 29, no. 11 (2019): 1805–1815, <https://doi.org/10.1101/gr.249805.119>.
244. L. Ren, M. Luo, J. Cui, et al., “Variation and Interaction of Distinct Subgenomes Contribute to Growth Diversity in Intergeneric Hybrid Fish,” *Genomics, Proteomics and Bioinformatics* 22 (2024): qzae055, <https://doi.org/10.1093/gpbjnl/qzae055>.
245. D. Gong, M. Tao, L. Xu, et al., “An Improved Hybrid Bream Derived From a Hybrid Lineage of *Megalobrama amblycephala* (♀) × *Culter alburnus* (♂),” *Science China Life Sciences* 65, no. 6 (2022): 1213–1221, <https://doi.org/10.1007/s11427-021-2005-5>.
246. S. Liu, S. Wang, Q. Liu, et al., “The Research Advances in Animal Distant Hybridization and Polyploid Organisms,” in *Fish Distant Hybridization* (Springer Nature Singapore, 2022), 1–37.
247. K. Zhang, X. Huang, C. Wang, et al., “Unveiling Potential Sex-Determining Genes and Sex-Specific Markers in Autotetraploid *Carassius auratus*,” *Science China Life Sciences* 67, no. 11 (2024): 2444–2458, <https://doi.org/10.1007/s11427-023-2694-5>.
248. A. Nagy, K. Rajki, J. Bakos, and V. Csányi, “Genetic Analysis in Carp (*Cyprinus carpio*) Using Gynogenesis,” *Heredity* 43 (1979): 35–40, <https://doi.org/10.1038/hdy.1979.57>.
249. C. E. Purdom, “Radiation-Induced Gynogenesis and Androgenesis in Fish,” *Heredity (Edinb)* 24, no. 3 (1969): 431–444, <https://doi.org/10.1038/hdy.1969.59>.



250. Y. Wang, A. M. Liao, H. Tan, et al., "The Comparative Studies on Growth Rate and Disease Resistance Between Improved Grass Carp and Common Grass Carp," *Aquaculture* 560 (2022): 738476, <https://doi.org/10.1016/j.aquaculture.2022.738476>.
251. S. Wang, Q. Xu, Q. Qin, et al., "The System Establishment of Improved Seed, Healthy Aquaculture and High-Quality Sales of Fish," *Journal of Fisheries of China* 47, no. 11 (2023): 119602–119625, <https://doi.org/10.11964/jfc.20230914174>.
252. S. Liu, J. Luo, J. Chai, et al., "Genomic Incompatibilities in the Diploid and Tetraploid Offspring of the Goldfish  $\times$  Common Carp Cross," *Proceedings of the National Academy of Sciences* 113, no. 5 (2016): 1327–1332, <https://doi.org/10.1073/pnas.1512955113>.
253. C. Wu, X. Huang, Q. Chen, et al., "The Formation of a New Type of Hybrid Culter Derived From a Hybrid Lineage of *Megalobrama amblycephala* ( $\text{♀}$ )  $\times$  *Culter alburnus* ( $\text{♂}$ )," *Aquaculture* 525 (2020): 735328, <https://doi.org/10.1016/j.aquaculture.2020.735328>.
254. P. Yu, H. Zhong, H. Chen, et al., "Study of Biological Characteristics of an Improved Japanese White Crucian Carp Lineage Derived From *Carassius cuvieri* ( $\text{♀}$ )  $\times$  *Megalobrama amblycephala* ( $\text{♂}$ )," *Aquaculture* 577 (2023): 739955, <https://doi.org/10.1016/j.aquaculture.2023.739955>.
255. W. He, L. Xie, T. Li, et al., "The Formation of Diploid and Triploid Hybrids of Female Grass Carp  $\times$  Male Blunt Snout Bream and Their 5S rDNA Analysis," *BMC Genetics* 14 (2013): 110, <https://doi.org/10.1186/1471-2156-14-110>.
256. M. Takeuchi, Y. Kawamura, T. Arai, et al., "The Size of the Sperm Head Influences the Gynogenetic Success in Teleost Fish," *Aquaculture* 596 (2025): 741768, <https://doi.org/10.1016/j.aquaculture.2024.741768>.
257. M. Wen, Y. Zhang, S. Wang, et al., "Characterization of Sex Locus and Sex-Specific Sequences in the Mandarin Fishes," *Aquaculture* 561 (2022): 738650, <https://doi.org/10.1016/j.aquaculture.2022.738650>.
258. M. Wen, S. Wang, C. Zhu, et al., "Identification of Sex Locus and a Male-Specific Marker in Blunt-Snout Bream (*Megalobrama amblycephala*) Using a Whole Genome Resequencing Method," *Aquaculture* 582 (2024): 740559, <https://doi.org/10.1016/j.aquaculture.2024.740559>.
259. M. Wen, Y. Zhang, S. Wang, et al., "Sex Locus and Sex Markers Identification Using Whole Genome Pool-Sequencing Approach in the Largemouth Bass (*Micropterus salmoides* L.)," *Aquaculture* 559 (2022): 738375, <https://doi.org/10.1016/j.aquaculture.2022.738375>.
260. S. Liu, M. Wen, X. Huang, et al., "The Formation and Biological Characteristics of the Different Ploidy Fishes Derived From the Hybridization of Red Crucian Carp  $\times$  Blunt Snout Bream," in *Fish Distant Hybridization* (Springer Nature Singapore, 2022), 145–187.
261. J. Wang, J. Xiao, M. Zeng, et al., "Genomic Variation in the Hybrids of White Crucian Carp and Red Crucian Carp: Evidence From Ribosomal DNA," *Science China. Life Sciences* 58, no. 6 (2015): 590–601, <https://doi.org/10.1007/s11427-015-4835-2>.
262. Q. Liu, J. Liu, Q. Liang, et al., "A Hybrid Lineage Derived From Hybridization of *Carassius Cuvieri* and *Carassius auratus red var.* and a New Type of Improved Fish Obtained by Back-Crossing," *Aquaculture* 505 (2019): 173–182, <https://doi.org/10.1016/j.aquaculture.2019.02.056>.
263. Q. Liu, K. Luo, X. Zhang, et al., "A New Type of Triploid Fish Derived From the Diploid Hybrid Crucian Carp ( $\text{♀}$ )  $\times$  Autotetraploid Fish ( $\text{♂}$ )," *Reproduction and Breeding* 1, no. 2 (2021): 122–127, <https://doi.org/10.1016/j.repbre.2021.07.003>.
264. X. Zhang, F. Liu, L. Bei, et al., "Comparative Analysis of Nutrients in Muscle and Ovary Between an Improved Fish and Its Parents," *Reproduction and Breeding* 4, no. 1 (2024): 16–21, <https://doi.org/10.1016/j.repbre.2023.12.002>.
265. N.-X. Xiong, Z.-X. Fang, X.-Y. Kuang, J. Ou, S.-W. Luo, and S.-J. Liu, "Integrated Analysis of Gene Expressions and Metabolite Features Unravel Immunometabolic Interplay in Hybrid Fish (*Carassius cuvieri*  $\text{♀}$   $\times$  *Carassius auratus Red Var*  $\text{♂}$ ) Infected With *Aeromonas hydrophila*," *Aquaculture* 563 (2023): 738981, <https://doi.org/10.1016/j.aquaculture.2022.738981>.
266. X. Zhang, F. Liu, B. Li, et al., "Multi-Omics Reveals the Molecular Mechanisms of Rapid Growth in Distant Hybrid Fish," *Aquaculture* 596 (2025): 741783, <https://doi.org/10.1016/j.aquaculture.2024.741783>.
267. Q. Liu, L. Duan, B. Li, et al., "The Key Role of Myostatin b in Somatic Growth in Fishes Derived From Distant Hybridization," *Science China. Life Sciences* 67, no. 7 (2024): 1441–1454, <https://doi.org/10.1007/s11427-023-2487-8>.
268. Q. Gu, S. Wang, H. Zhong, et al., "Phylogeographic Relationships and the Evolutionary History of the *Carassius auratus* Complex With a Newly Born Homodiploid Raw Fish (2nNCRC)," *BMC Genomics* 23, no. 1 (2022): 242, <https://doi.org/10.1186/s12864-022-08468-x>.
269. S. Fan, Z. Tang, Y. Wang, et al., "The Bisexual Natural Gynogenetic Blunt Snout Bream Lineage Derived From the Distant Hybridization of Female Blunt Snout Bream and Male Bleeker's Yellow Tail," *Aquaculture Reports* 37 (2024): 102206, <https://doi.org/10.1016/j.aqrep.2024.102206>.
270. H. Tan, B. Hu, W. Liu, et al., "Comprehensive Analysis of the Immunological Differences in the Intestinal Barrier of Improved Grass Carp and Their Parents," *Aquaculture* 577 (2023): 739931, <https://doi.org/10.1016/j.aquaculture.2023.739931>.

## Biographies

**Qizhi Liu** focuses on distant hybridization and gynogenesis of *Siniperca chuatsi*, innate immunity of Hefang crucian carp, and splicing coupling transcription in human and fish cells. He has published 19 SCI paper.

**Anmin Liao** mainly engages in research on fish genetics and breeding, focusing on distant hybridization of fish and gynogenesis in fish. He has achieved innovative research results in the creation of gynogenetic fish strains.

**Min Tao** is the Young Scholars of the Cheung Kong Scholars Award Program, Leading talents in science and technology innovation in Hunan Province. Research interest is fish genetics and breeding research. She has presided over a number of national, provincial projects. More than 30 papers were published in SCI.

**Qinbo Qin** mainly engaged in the research of fish genetic breeding and healthy breeding. Many kinds of fish with excellent traits, such as autotetraploid fish and its all-female autotetraploid gynogenetic offspring, have been prepared by distant hybridization, gynogenesis and androgenesis.

**Kaikun Luo** has been working on distant hybridization of fish for more than 30 years. He created fish lineages including Hefan No. 2 crucian carp, Hefan crucian carp, and *Culter alburnus*. He has won second prizes of the China National Science and Technology Progress Award twice.

**Chun Zhang**, a researcher (Professor) at Hunan Normal University in China. She is a project leader of the National Key R&D Program of China and has been a visiting scholar at Stockholm University in Sweden, mainly engaging in fish genetics and breeding.

**Shi Wang** is mainly engaged in the research of fish genetics and breeding, focusing on the phenomenon of genome collision between homoploid and polyploid offspring of distant hybridization in fish, and has made innovative research achievements in the creation of autotetraploid and autotetraploid fish lineages derived from distant hybridization of fish.

**Yi Zhou** is an expert in the field of fish genetics and breeding. With over 20 years of experience in the field, he has dedicated his career to advancing fish germplasm by hybridization. His work focuses on enhancing genetic diversity, growth rate, and meat quality in farmed fish populations.

**Fangzhou Hu** have accumulated extensive research experience in the fields of fish genetic breeding and related fields. The current research focuses on the improvement of freshwater fish germplasm by using traditional breeding methods (hybridization and gynogenesis), and explore of genes involved in superiority traits in hybrids or gynogenetic fish.



**Yude Wang**, male, Doctor, Associate professor, master mentor. He mainly engaged in fish genetic breeding and seed industry research. he has two national key research and development projects, more than 20 invention papers, 10 patents approved, and participated in the development of high-quality fish in a number of provinces and cities for large-scale farming.

**Chang Wu** has been engaged in fish genetics and breeding for more than 10 years, focoused on the genetic improvement of culter and the research of hybrid lineages of blunt black bream and topmouth culter, and created a series of new hybrid lineages and gynogenetic population by distant hybridization and gynogenesis technology.

**Wuhui Li** primarily focuses on fish genetic breeding and ecological health aquaculture research. As the first author, he has published 14 papers in journals such as Genome Research and Aquaculture.

**Qingfeng Liu** is working on fish genetics and breeding. His major research interest is the genetic mechanisms analysis of hybrid fish dominant traits. He has published over 30 articles.

**Chenchen Tang** is working as a research technician focusing on the genetic breeding of fish and secretary of Hunan Fisheries Society. He won the Science and Technology Innovation Team Award of Hunan Province in 2021.

**Jing Wang** is using distant hybridization to provide further insight into the mechanism of sterility and the formation of unreduced gametes. The study provides a framework for the further exploitation and application of hybrid germplasms.

**Rurong Zhao** has led a project funded by the National Natural Science Foundation of China's Youth Science Fund, focusing on the identification of the Kisspeptin/GPR54 signaling system in infertile female triploid fish and its role in regulating the HPG axis. She has also led a key research and development project in Hunan Province on the health enhancement, aquaculture technology, genetic improvement, and industrialization of Topmouth Culter

**Shaojun Liu** leads a team that has been engaged in fish genetic breeding research for a long time. He has overcome the challenge of reproductive barriers and explored the main genetic and reproductive rules of distant hybridization in fish. He established key breeding techniques for fish hybridization using both one-step and multi-step methods.